

# Cancer Biology 2

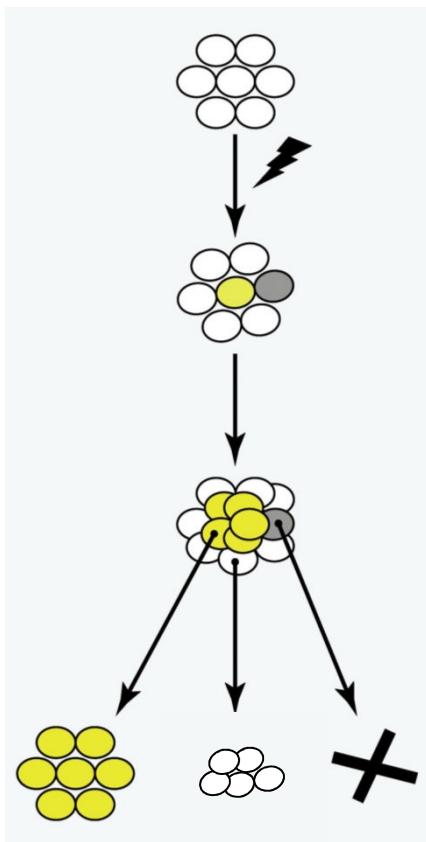
Cancer Biology 2			
<b>week</b>	<b>date</b>	<b>teacher</b>	<b>topic</b>
<b>1</b>	2025-02-17	Joerg Huelsken	Cancer Stem Cells
<b>2</b>	2025-02-24	Joerg Huelsken	Metastasis
<b>3</b>	2025-03-03	Joerg Huelsken	Cell death
<b>4</b>	2025-03-10	Joerg Huelsken	Cancer Signalling Pathways
<b>5</b>	2025-03-17	Joerg Huelsken	Tumor Histology
<b>6</b>	2025-03-24	Joerg Huelsken	Tumor Histology
<b>7</b>	2025-03-31	Joerg Huelsken	exam
<b>8</b>	2025-04-07	Joerg Huelsken	Adaptive Immunity (T cells)
<b>9</b>	2025-04-14	Miki de Palma	Tumor Angiogenesis
<b>10</b>	2025-04-28	Miki de Palma	Tumor Angiogenesis
<b>11</b>	2025-05-05	Joerg Huelsken	Innate Immunity (Myeloid)
<b>12</b>	2025-05-12	Joerg Huelsken	Innate Immunity (NK cells)
<b>13</b>	2025-05-19	Joerg Huelsken	Cancer Metabolism
<b>14</b>	2025-05-26	Joerg Huelsken	exam

## Learning objectives

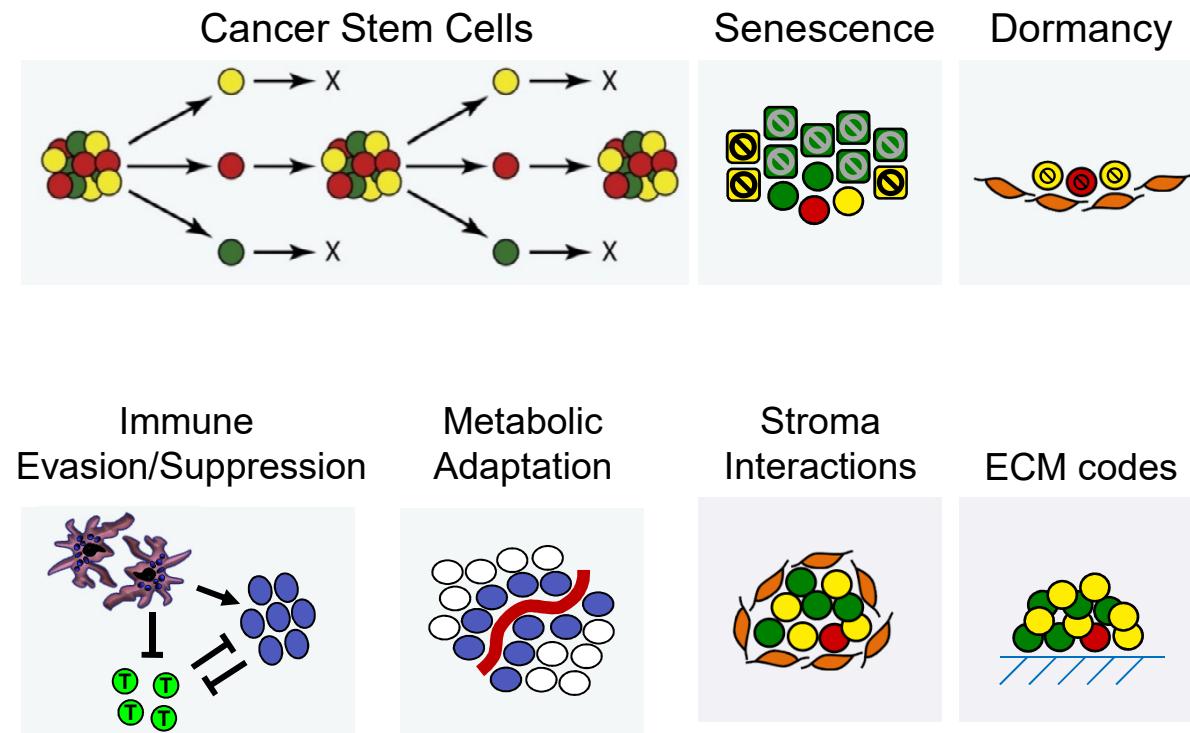
- Hierarchical organization in normal tissues and cancer
- Definition and functional properties of Cancer Stem Cells (CSCs)
- Assays and methods to detect CSCs
- In vivo evidence for CSCs
- Prospects of Targeting CSCs

# Intra-tumoral Heterogeneity

## Genetic Evolution

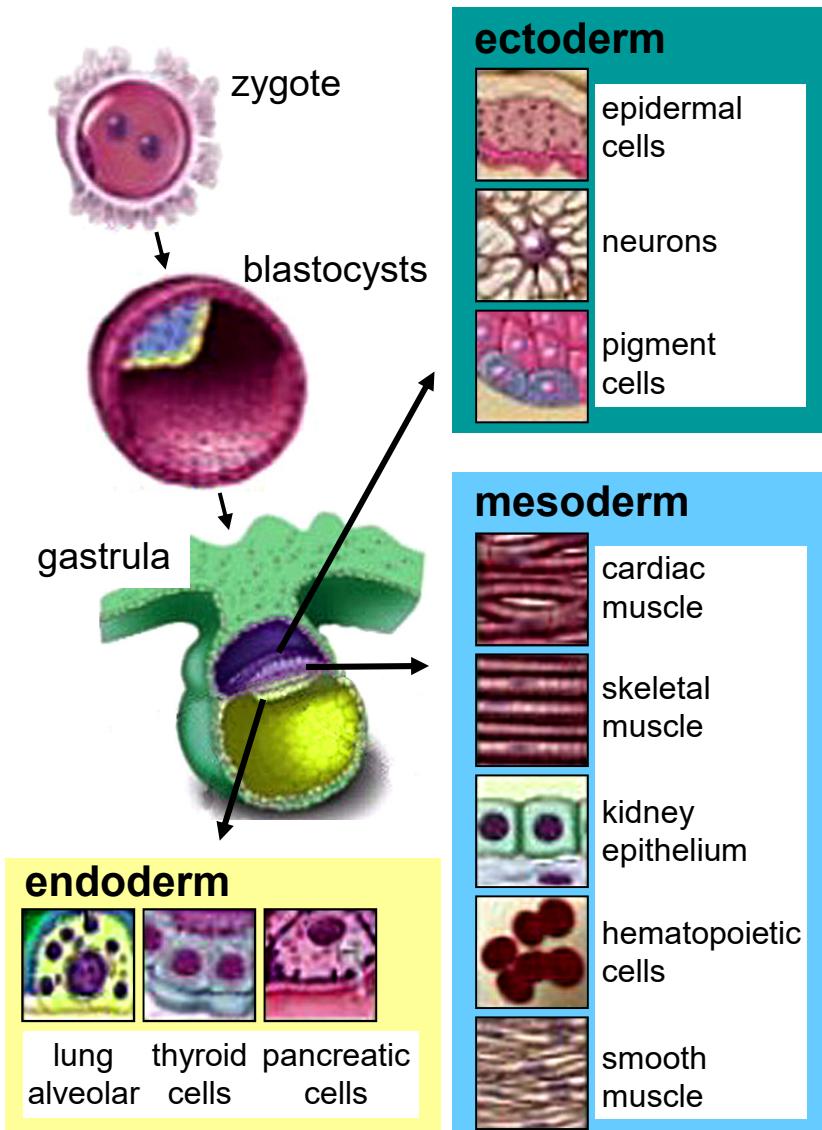
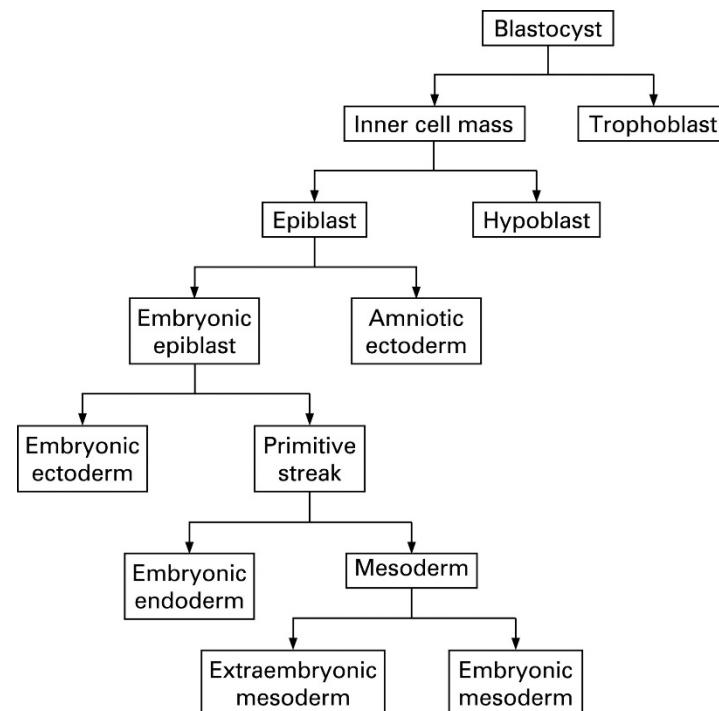


## Epigenetic and Environmental Adaptations

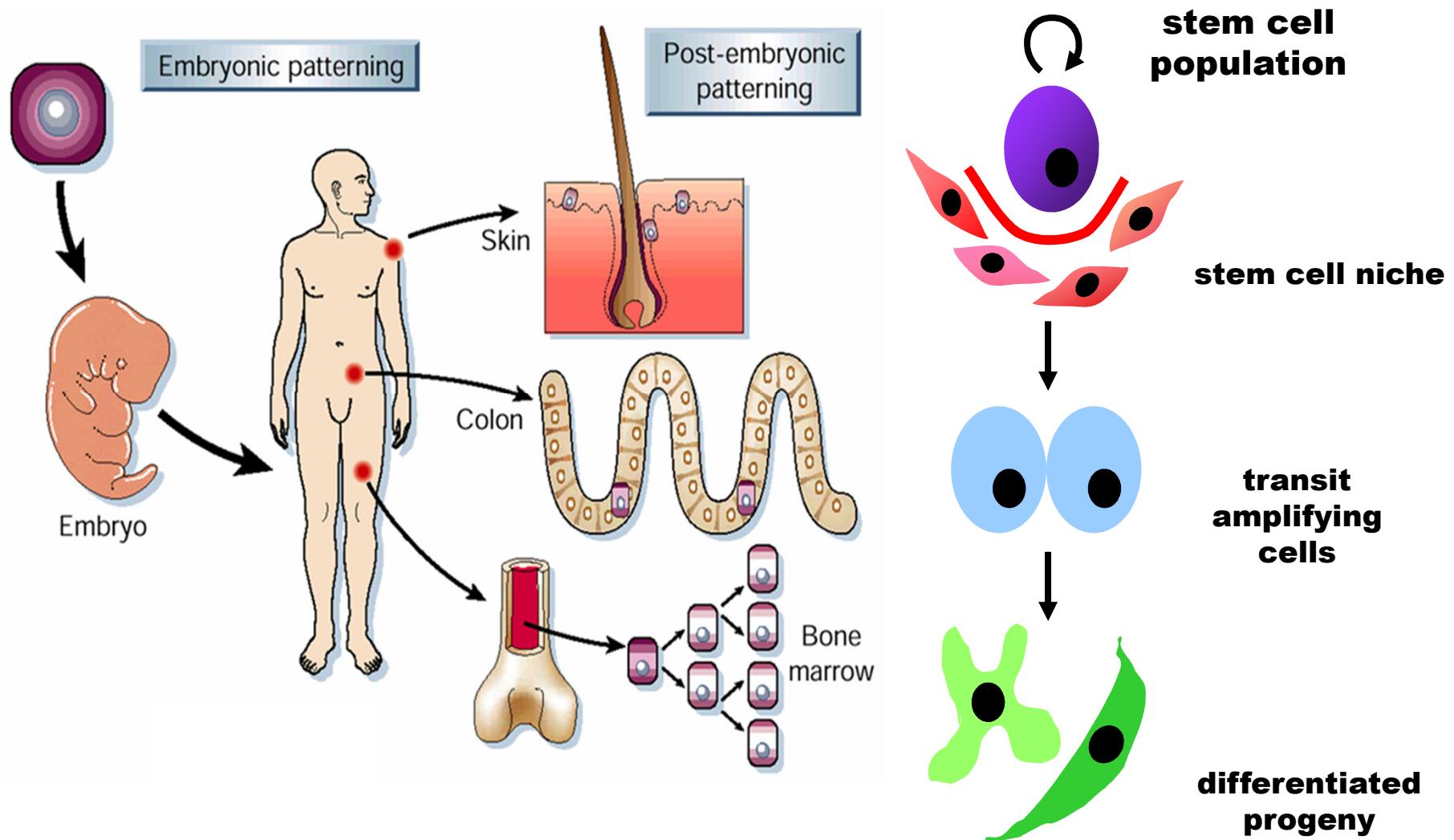


# Hierarchical organization principles in development

- hierarchical organization
- transient progenitor populations with increasingly narrowed potential
- successive patterning and specification events
- repeated use of few, conserved signaling pathways (Wnt, BMP, FGF, HH, Notch, ...)



# Hierarchical organization in tissue homeostasis



# CSC in AML: the first study

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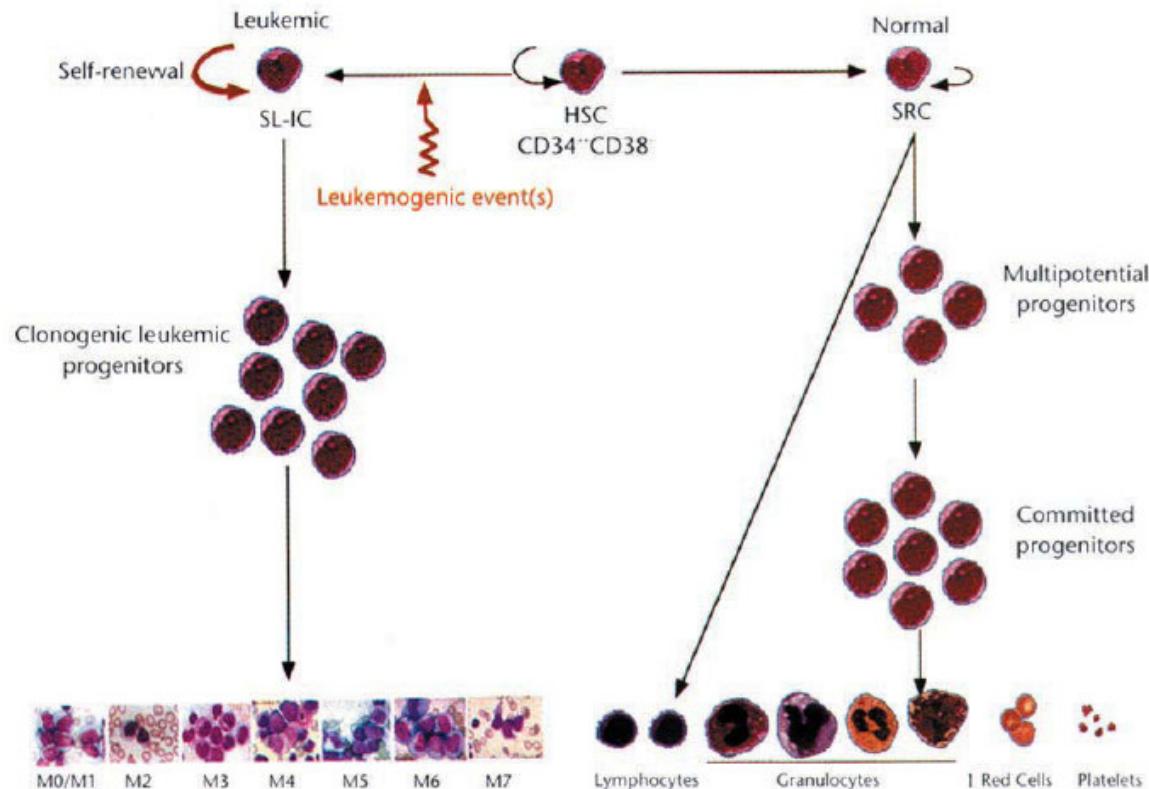
Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell

DOMINIQUE BONNET & JOHN E. DICK

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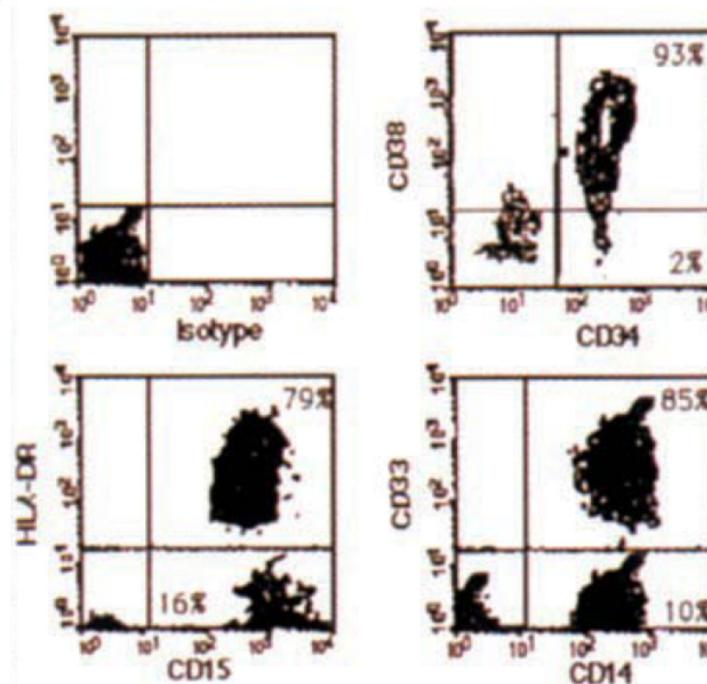
# CSC in AML: the first study

## frequency of CSCs and their multipotency

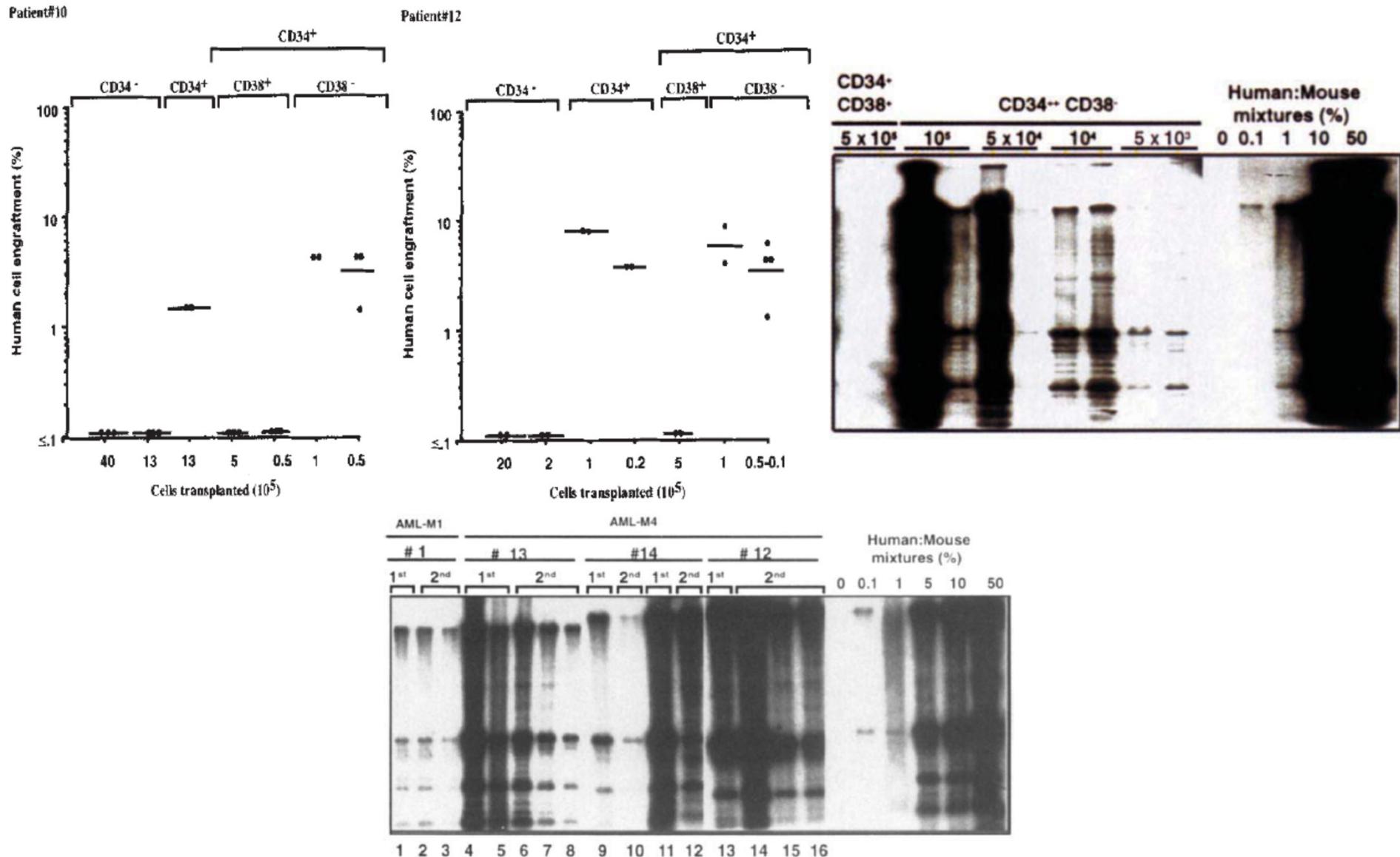
Table 1 Patient-to-patient heterogeneity in expression of CD34 and CD38 antigens

Patient	FAB subtype	Age/Sex	Level of engraftment of NOD/SCID mice with 10–20 × 10 <sup>6</sup> MNCs	Percent of CD34 <sup>+</sup> in MNCs	Percent of CD34 <sup>+</sup> CD38 <sup>+</sup> in MNCs	Estimated frequency of SL-IC per 10 <sup>6</sup> MNCs
1	M1	64/F	74 ± 10	43	0.8	100–200
8	M4	62/F	45 ± 8	80	1.0	1
10	M4	58/M	62 ± 5	11	0.75	0.2
12	M4	65/M	76 ± 6	2.0	0.2	49
13	M4	69/M	37 ± 7	95	2.0	0.2
14	M4	59/F	28 ± 9	1.1	0.2	2
18	M5	71/F	18 ± 6	0.3	0.02	0.2

FAB, French-American-British criteria<sup>6</sup> for subtypes; NOD/SCID mice, non-obese diabetic mice with severe combined immunodeficiency disease; MNCs, mononuclear cells; SL-IC, SCID leukemia-initiating cell.



# CSC in AML: the first study serial transplantation assays

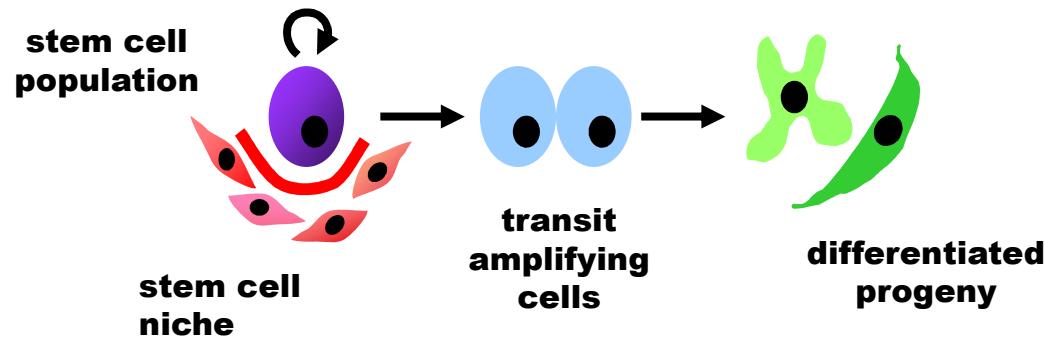


## Definition of **Tissue-specific** vs. **Cancer** Stem Cells

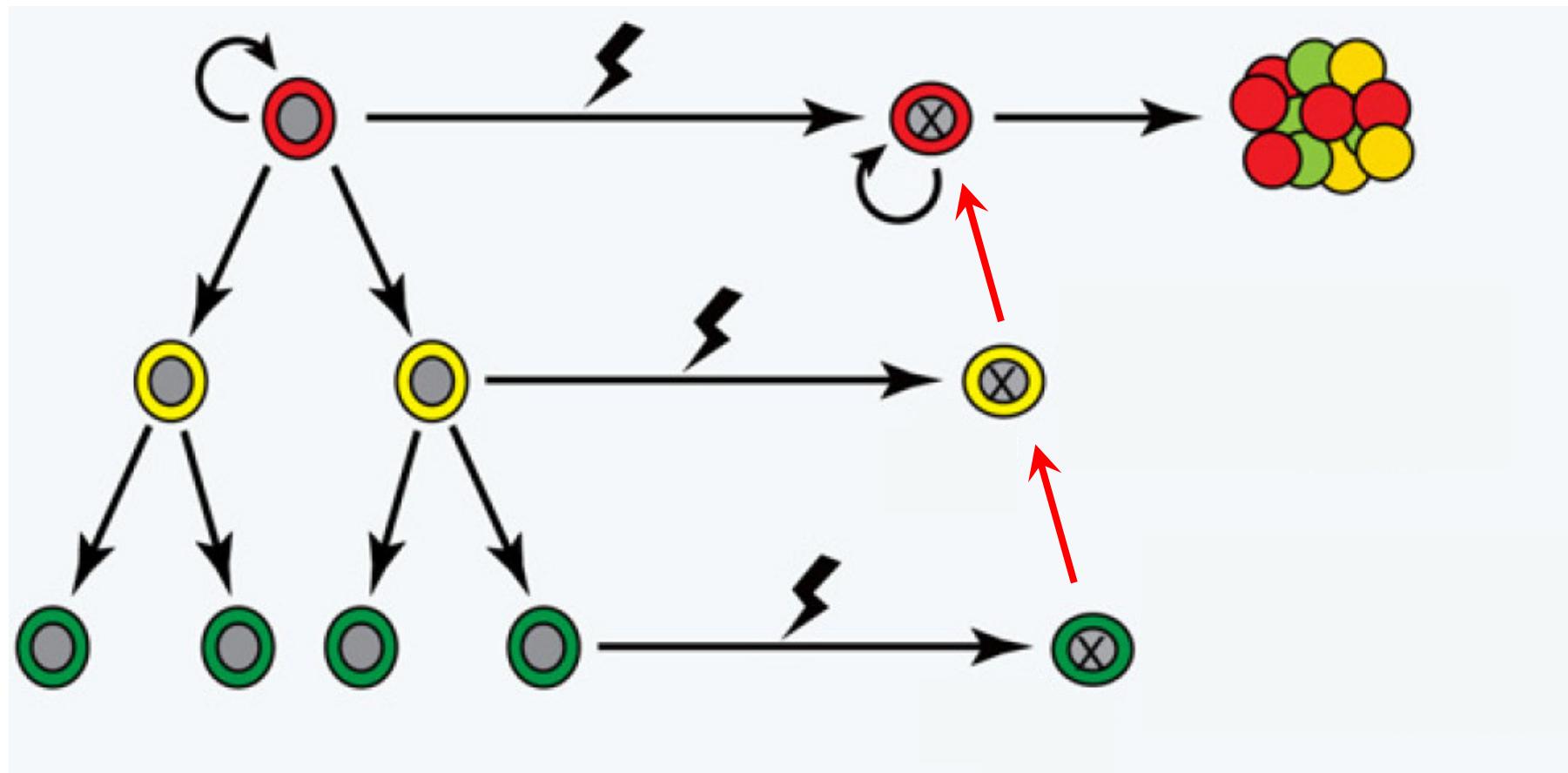
heterogeneous population of undifferentiated cells,

defined by functional assays and capable of:

- 1) self-renewing/self-maintaining their population  
self-renewing/self-maintaining their population
- 2) production of differentiated, functional progeny (multipotency)  
production of heterogeneic, (abberantly) differentiated progeny
- 3) regenerating a functional tissue after injury or upon transplantation  
regenerating an exact copy of the primary tumor upon transplantation
- 4) interaction with an appropriate environment (niche), homing to the niche  
self-sufficiency in growth stimulatory signals, evasion of growth-restrictive signal

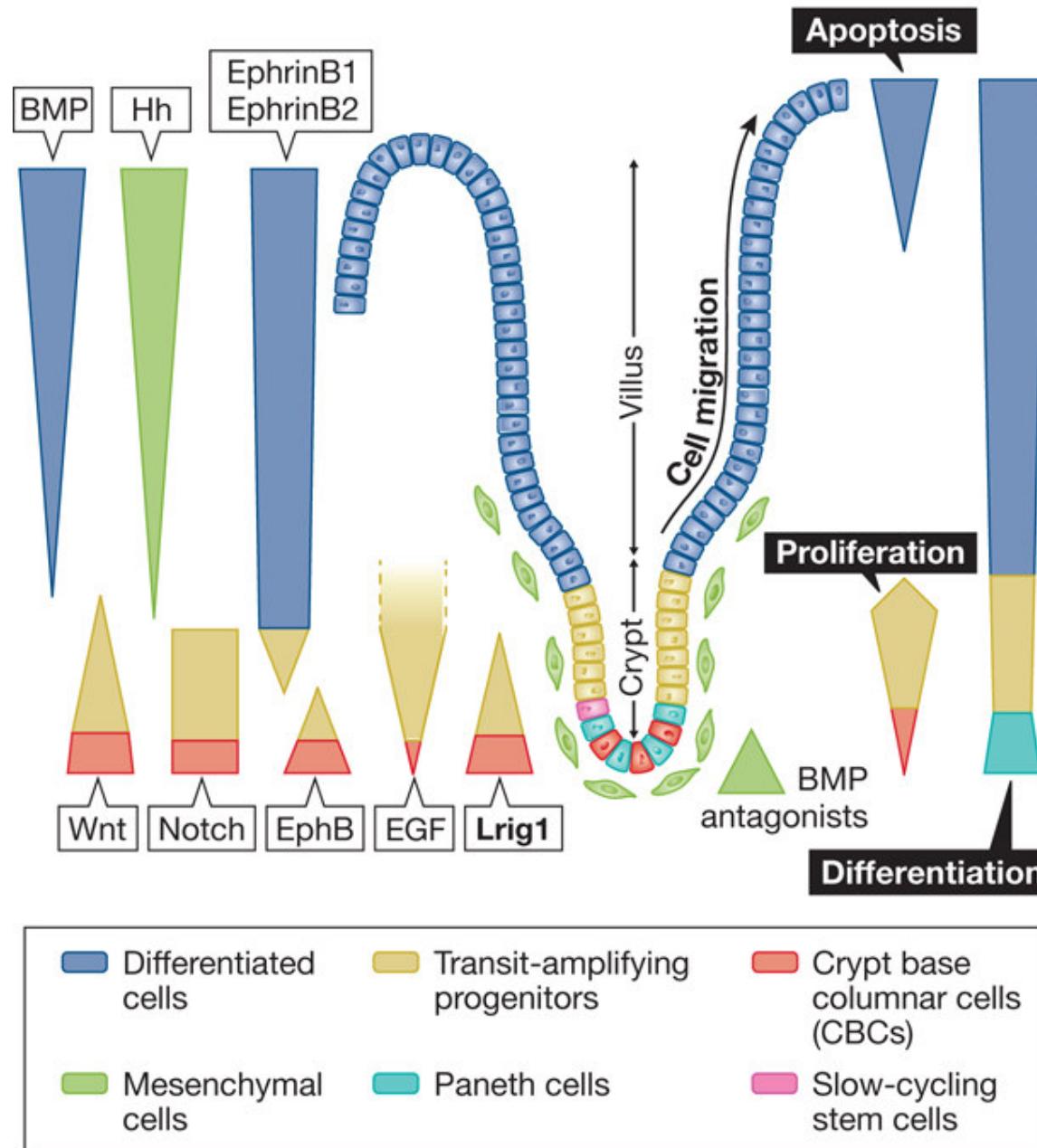


# Cell of origin

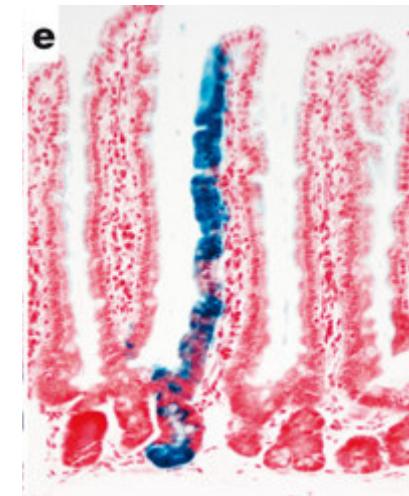
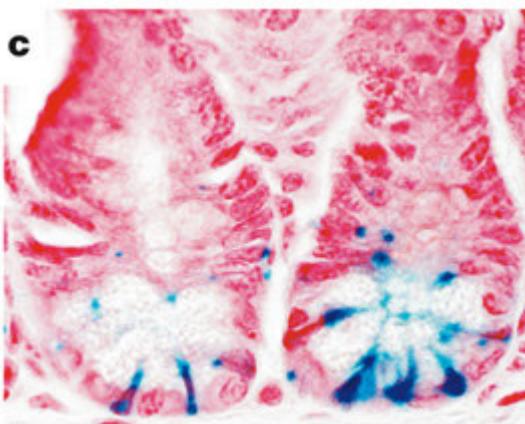
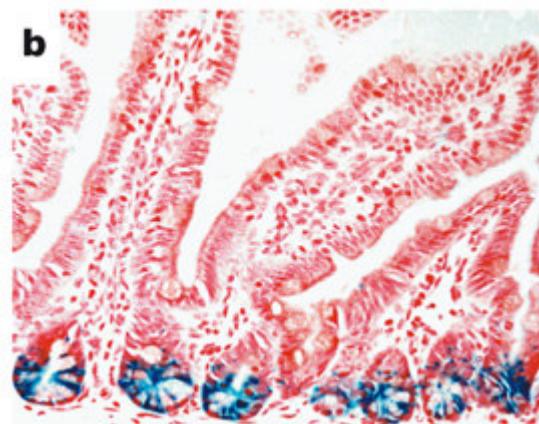
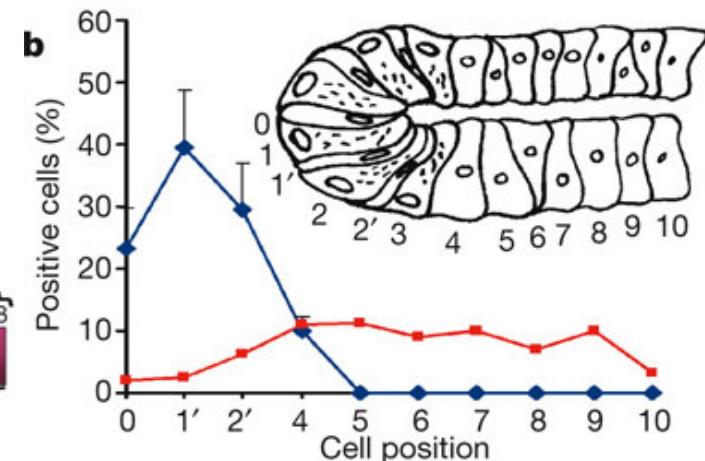
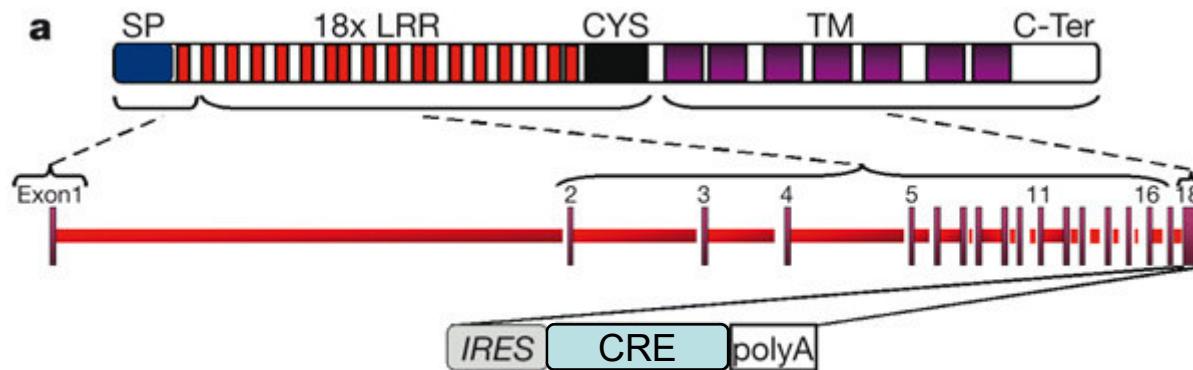


modified from Magee et al. *Cancer Cell* 2012

# Cues controlling intestinal homeostasis

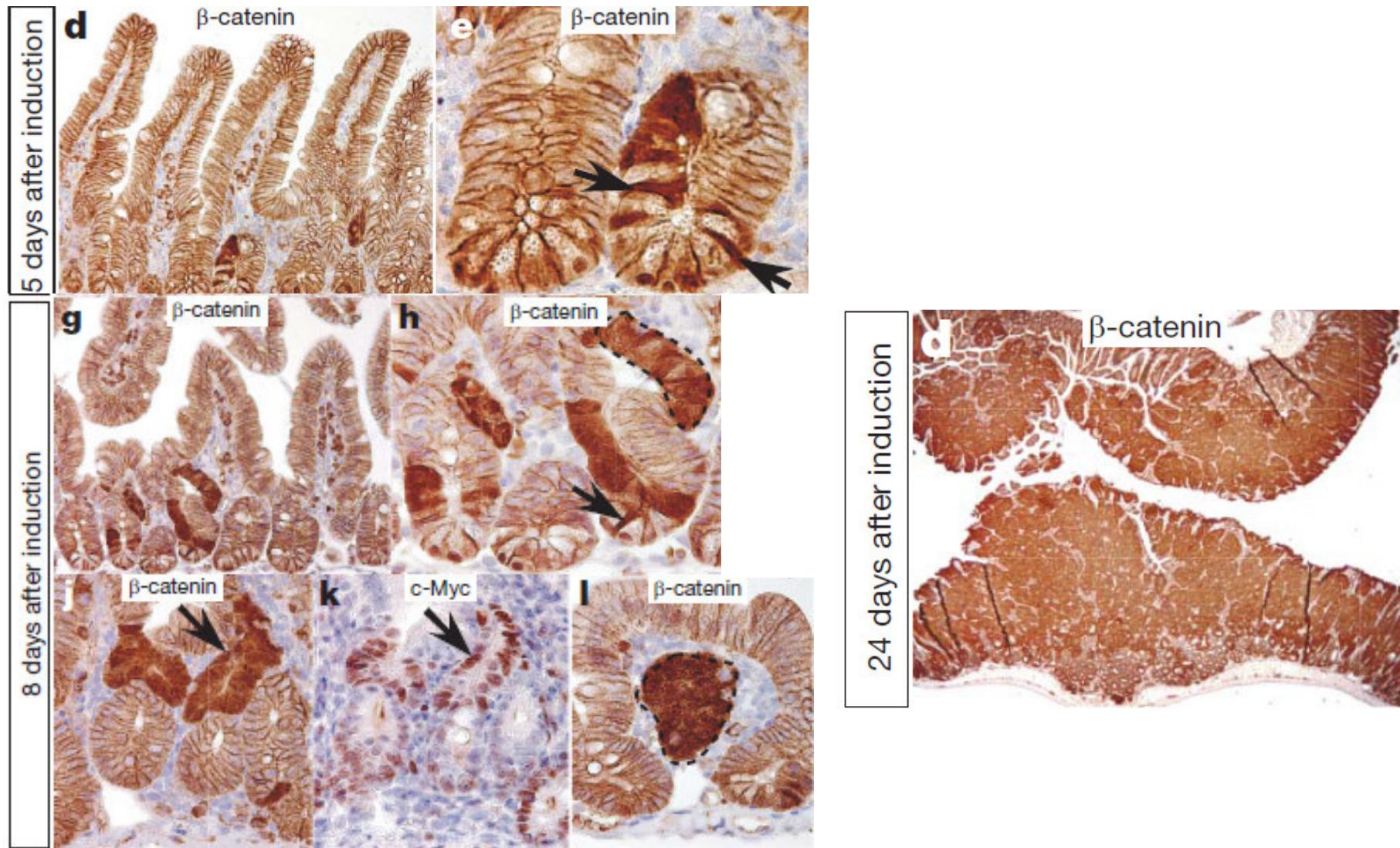


# Intestinal stem cell specific gene modification (Lgr5-Cre)

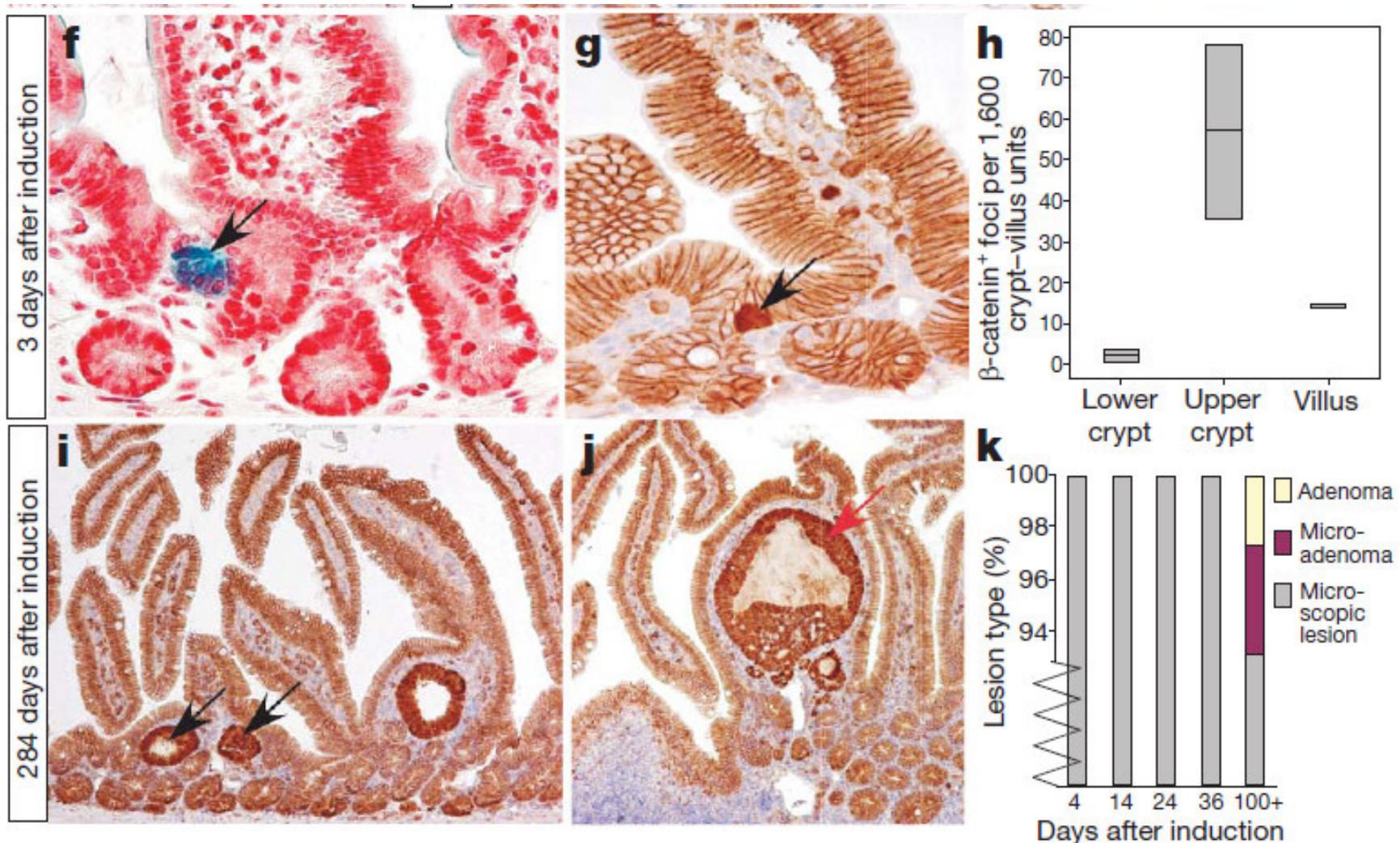


2 months

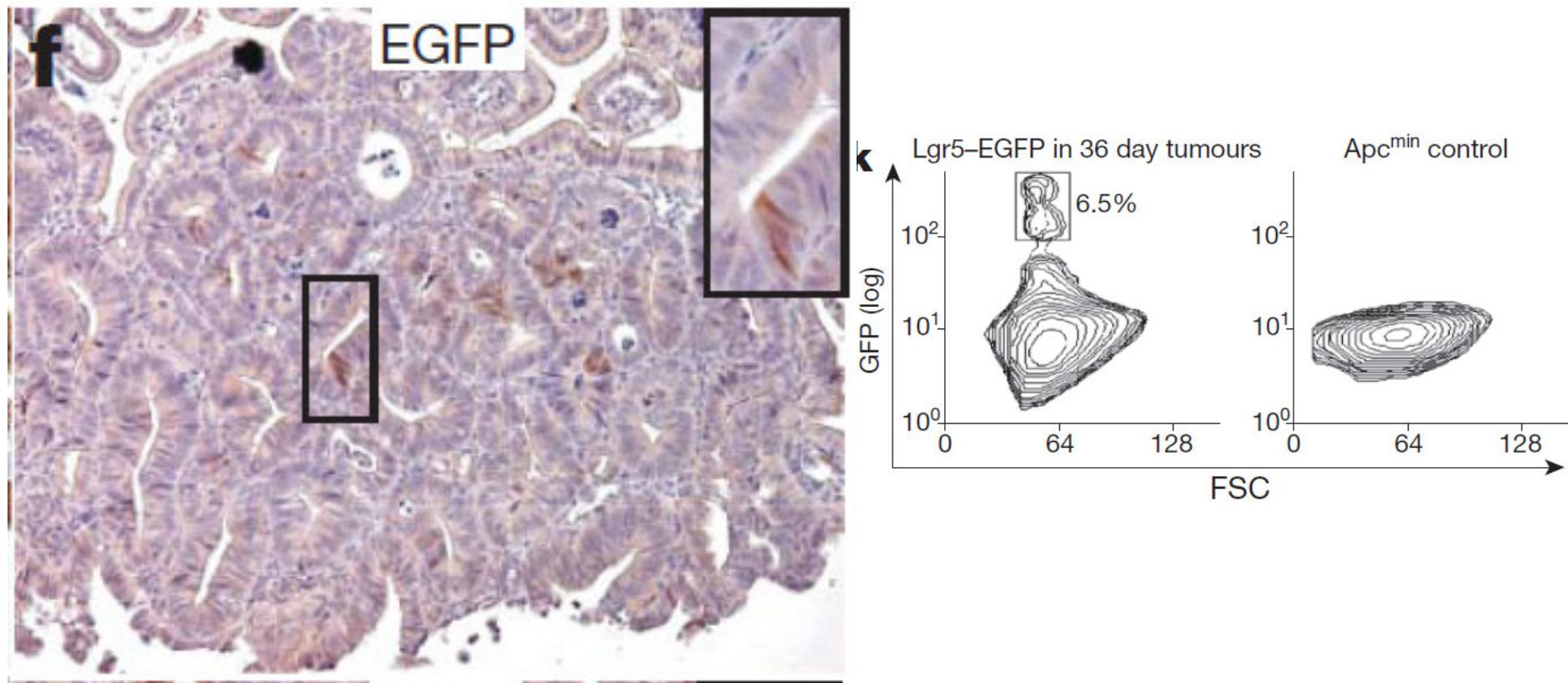
# Stem cell specific induction of APC loss induces cancer (Lgr5-Cre x APC<sup>lox/lox</sup>)



# APC loss in any epithelial cell fails to induce cancer (Ah-Cre x APC $^{lox/lox}$ )

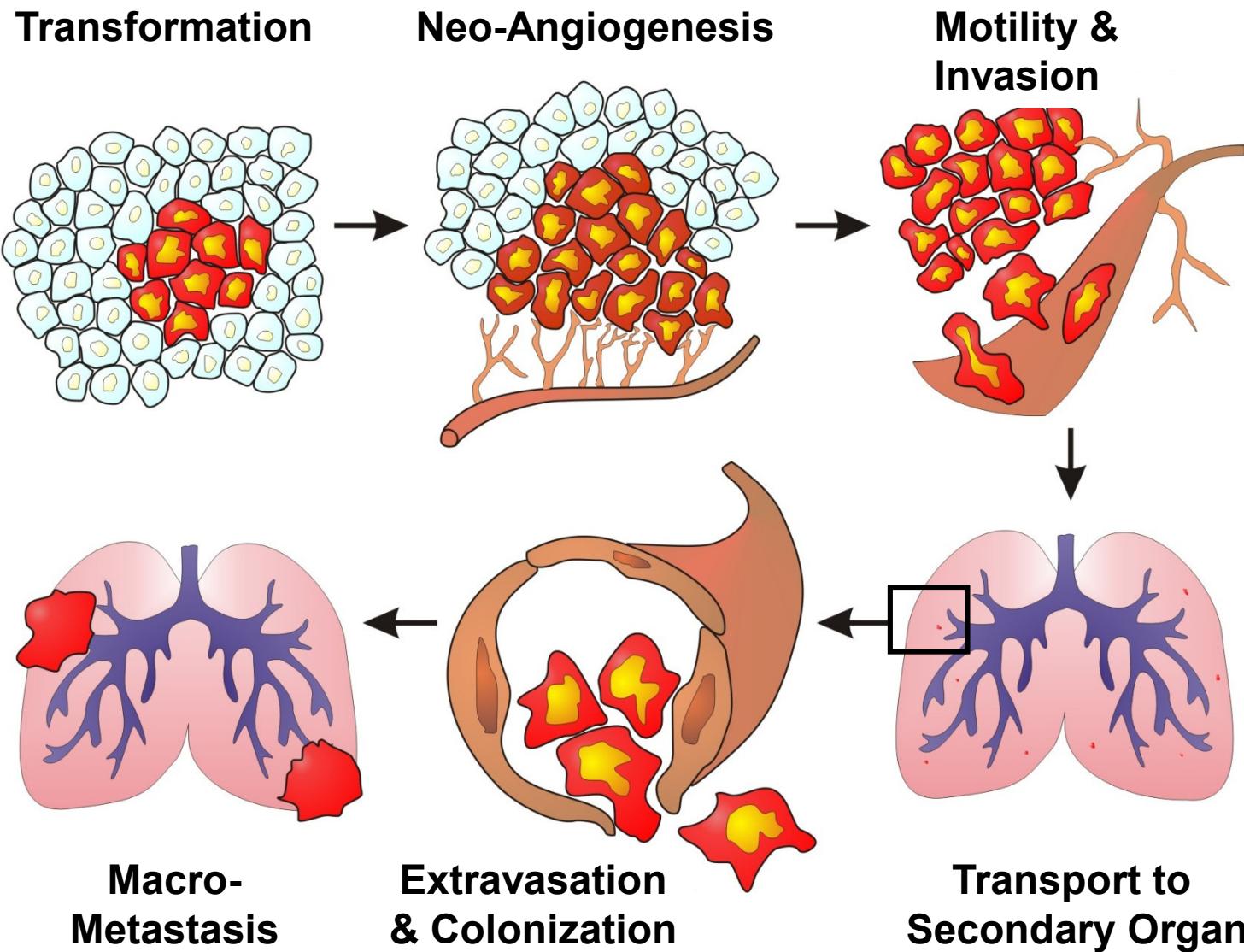


## Rare Lgr5+ population in adenomas



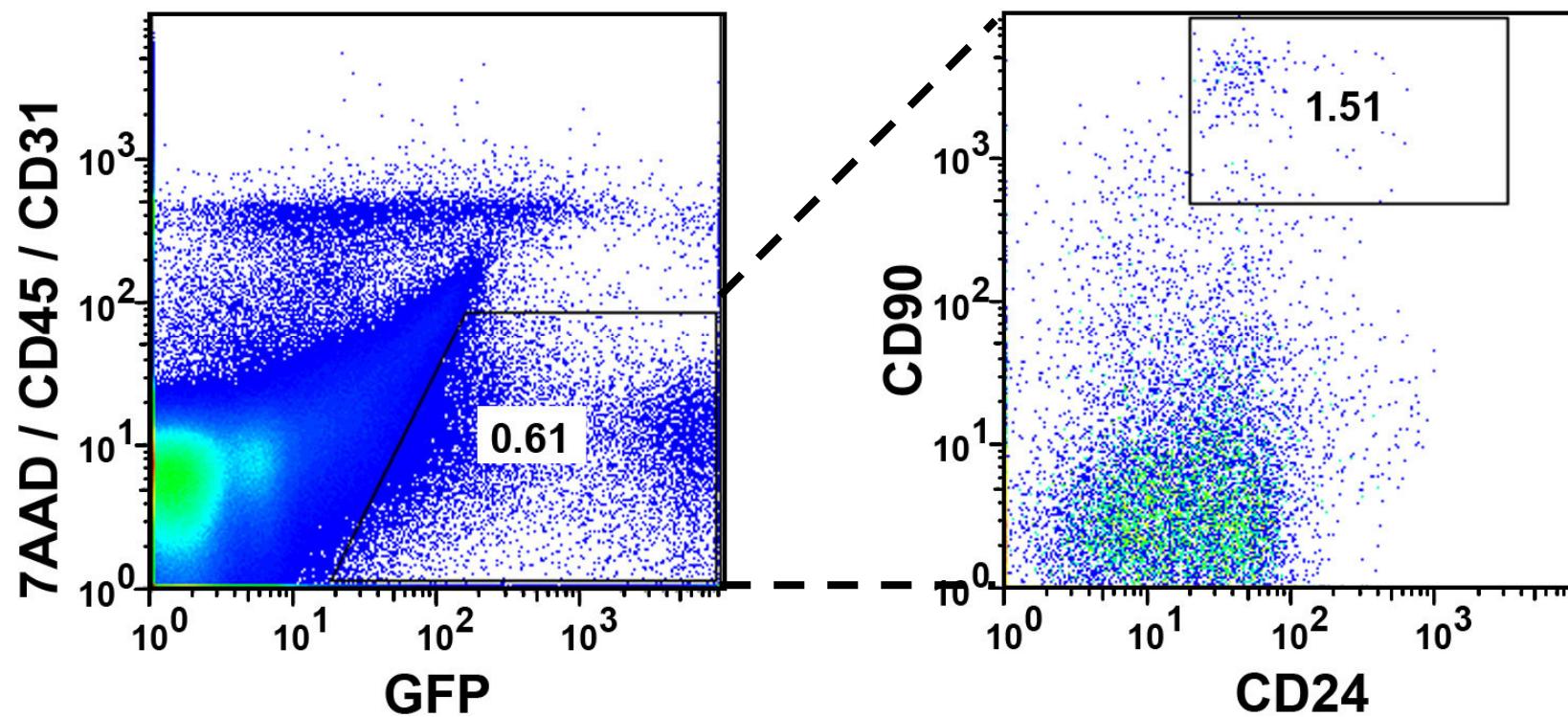
## **CSCs in cancer progression**

# Linear Cancer Progression Model



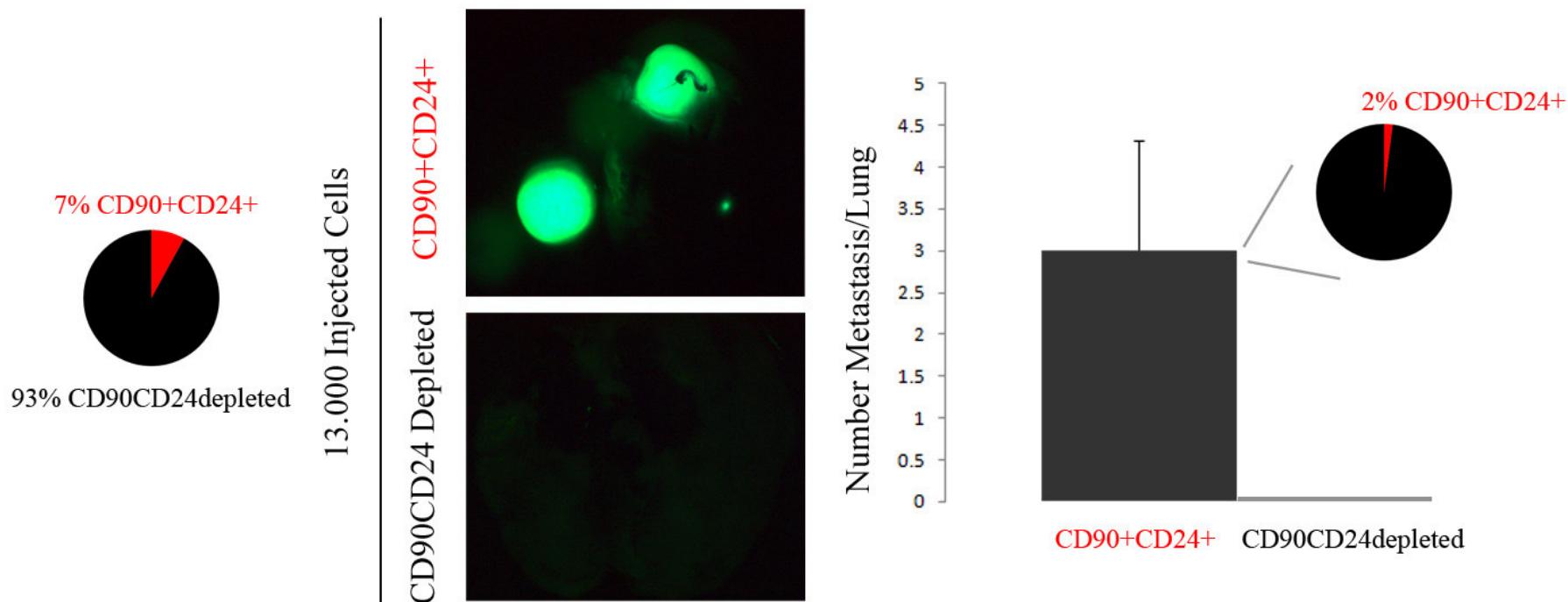
# Murine breast cancer model: MMTV-PyMT

## Isolation of CD24<sup>+</sup>CD90<sup>+</sup> cancer stem cells



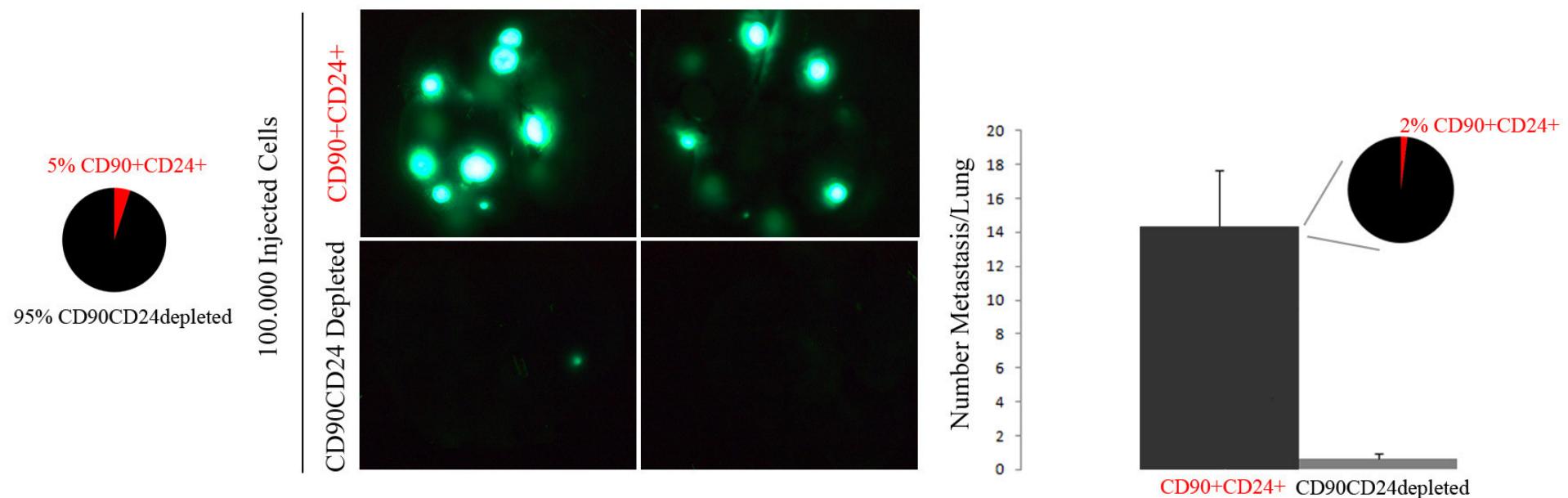
# The CD24<sup>+</sup>CD90<sup>+</sup> population is responsible for metastasis initiation

(cells isolated from primary tumors)

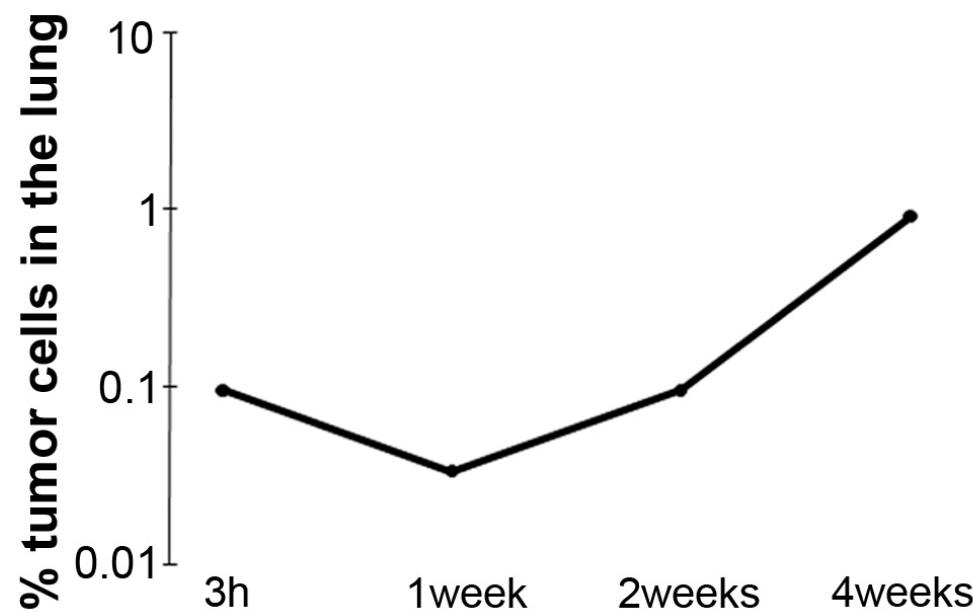
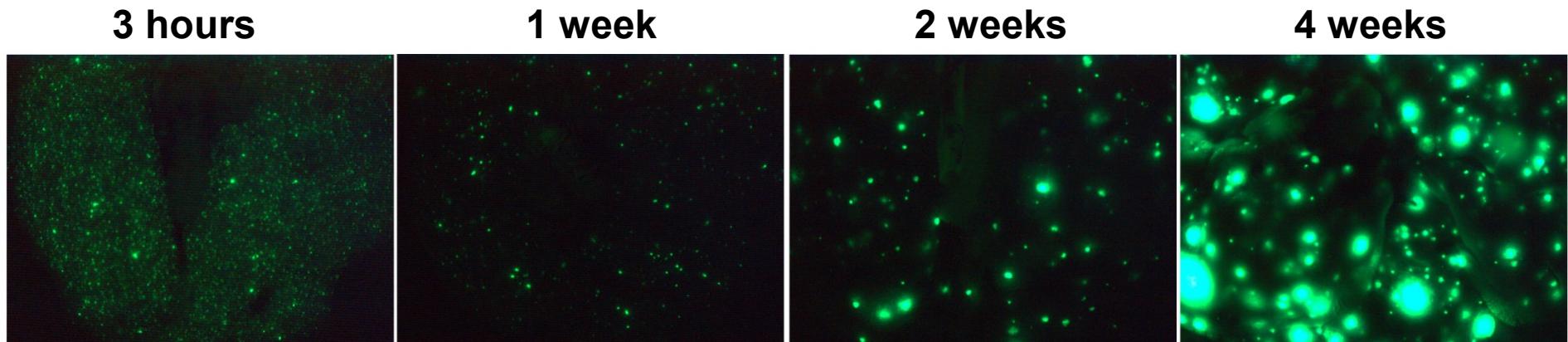


# The CD24<sup>+</sup>CD90<sup>+</sup> population is responsible for metastasis re-initiation

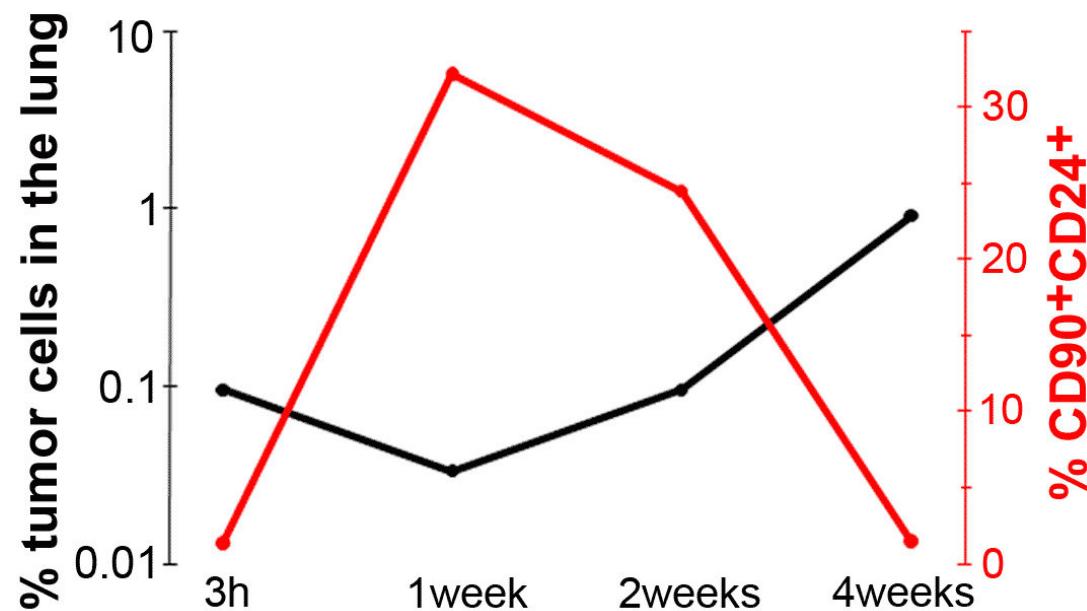
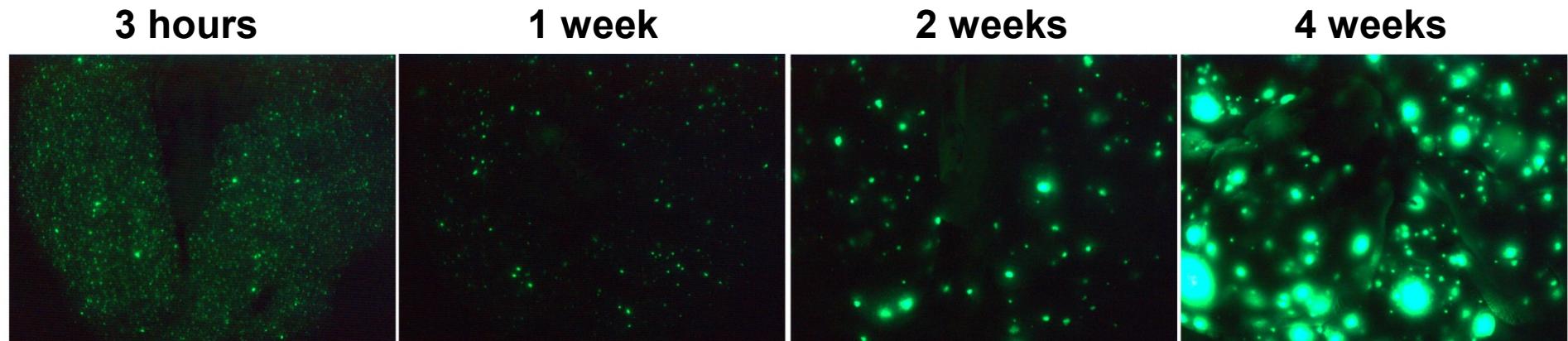
(cells isolated from metastasis)



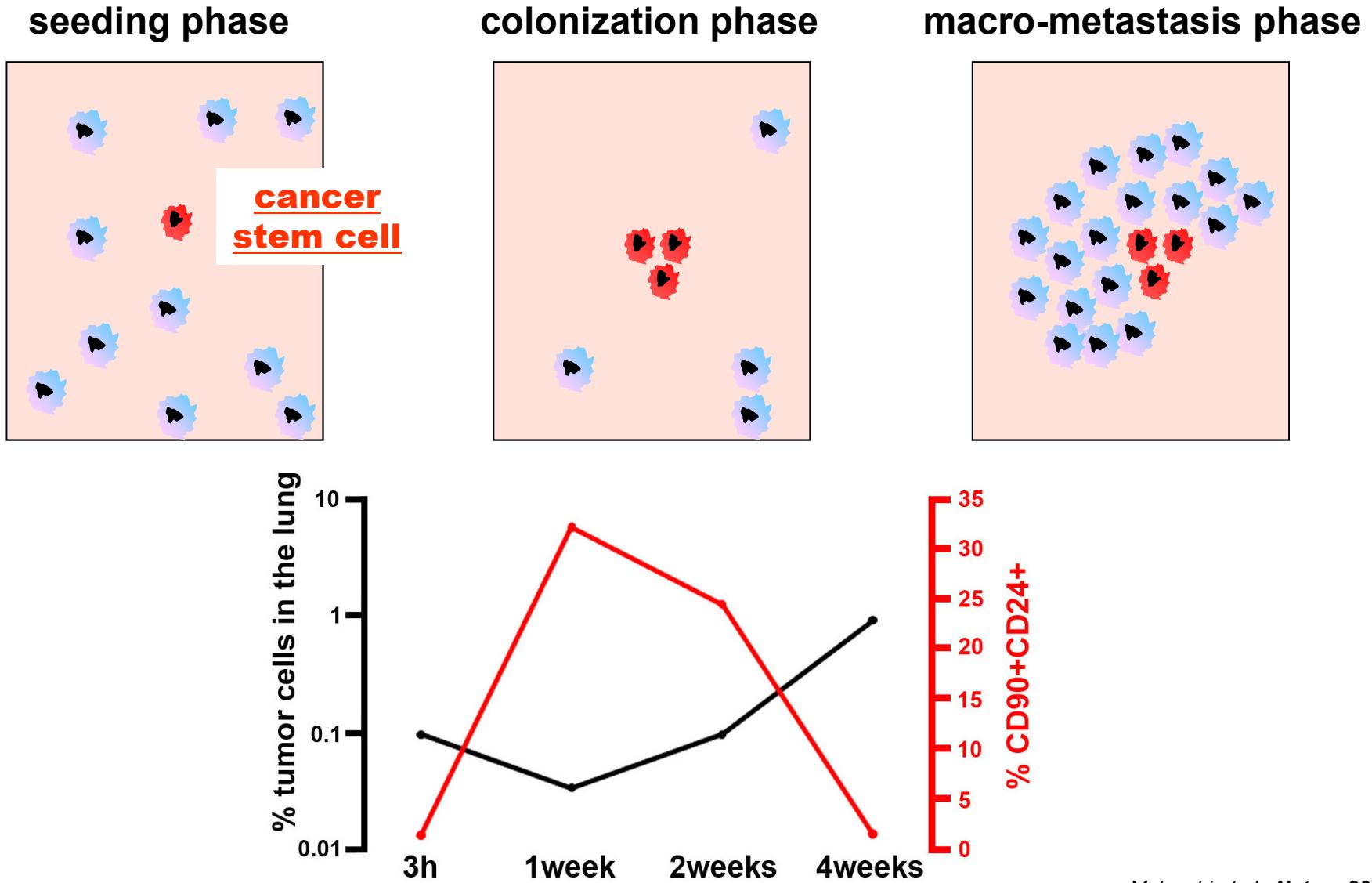
# Early metastatic colonization relies on cancer stem cells



# Early metastatic colonization relies on cancer stem cells



# Changes in the Relative Amount of Cancer Stem Cells during Metastatic Progression



# Functional characterization of CSCs

# Assaying CSCs: Transplantation

## - Transplantation

the assays to functionally identify a CSC population require transplantation into mice

- **which recipient strain?**

immuno-deficient for xenotransplantation of human cells or mouse cells with foreign cDNAs

- *Nude* (no thymus => hardly any  $\alpha\beta T$  cells, but B cells,  $\gamma\delta T$  cells and all other innate immune cells)
- *RAG* or *NOD/SCID* (neither T nor B cells, but innate immune cells)
- *RAG/cg* or *NOD/SCID/cg=NSG* (in addition lack of NK cells, less myeloid cells)

- **which route?**

- subcutaneous (artificial morphology)
- kidney capsule
- orthotopic (good morphology and invasion possible, but often difficult to perform)
- tail vein (metastasis-type of experiments, requires high cell numbers)

- **which other variables?**

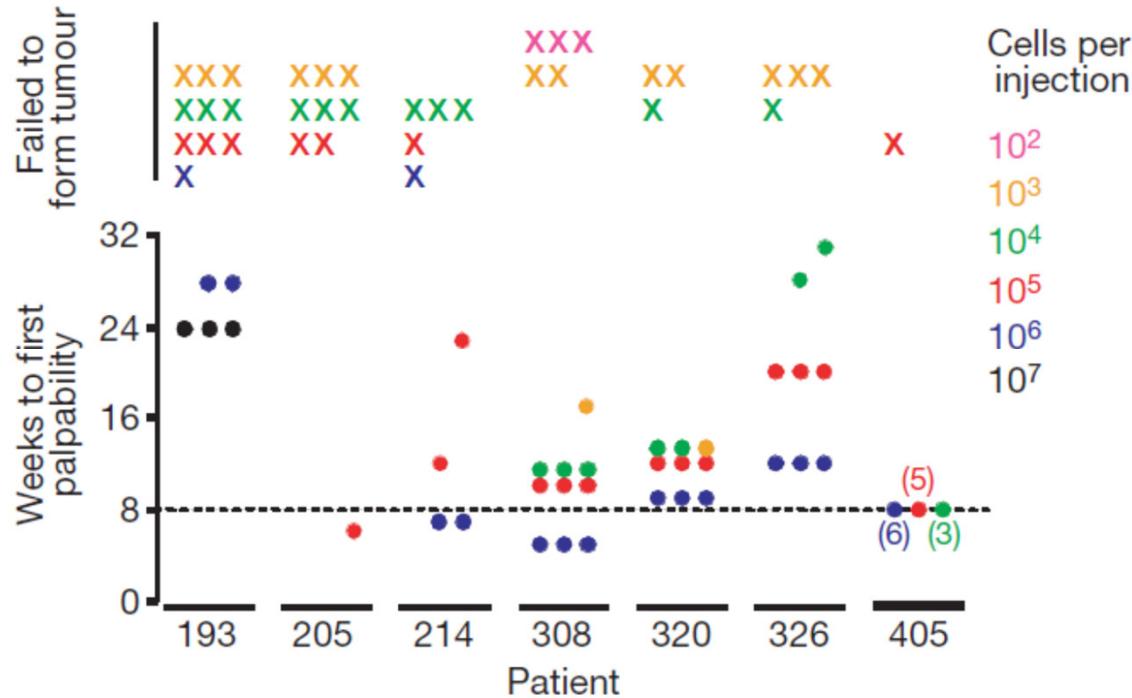
- cells embedded in ECM proteins

- **only selection for the most robust cells?**

- **only selection for cells which can cope with the species barrier (lack of factors)?**

# Detection of CSCs can depend on the transplantation system

transplantation into *NOD/SCID*



Time after injection	Melanoma-initiating cell frequency (95% confidence interval)
8 weeks	1/837,000 (1/512,000–1/1,370,000)
32 weeks	1/111,000 * (1/67,000–1/185,000)

# Detection of CSCs can depend on the transplantation system

transplantation into NSG

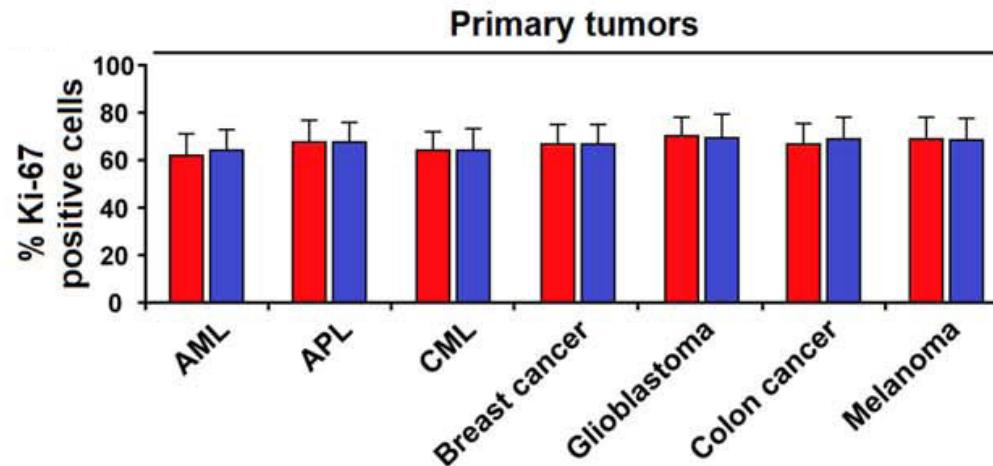
Patient	Mouse strain	Co-injection	Number of tumours / number of injections				Melanoma-initiating cell frequency (95% confidence interval)	
			cells per injection					
			50,000	5,000	50	5		
481	NOD/SCID	Vehicle	0/3	0/6	0/3		(<1/60,000)	
	NOD/SCID <i>Il2rg</i> <sup>-/-</sup>	Matrigel			6/6	4/6	1/5*	
491	NOD/SCID	Vehicle		0/3	0/6		(<1/5,100)	
	NOD/SCID <i>Il2rg</i> <sup>-/-</sup>	Matrigel			6/6	1/6	1/15*	
492	NOD/SCID	Vehicle	3/3	3/6	0/6		1/7,300	
	NOD/SCID <i>Il2rg</i> <sup>-/-</sup>	Matrigel			6/6	2/6	1/11*	
All	NOD/SCID	Vehicle	3/6	3/15	0/15		1/46,700	
	NOD/SCID <i>Il2rg</i> <sup>-/-</sup>	Matrigel			18/18	7/18	1/9*	

## ARTICLES

### Efficient tumour formation by single human melanoma cells

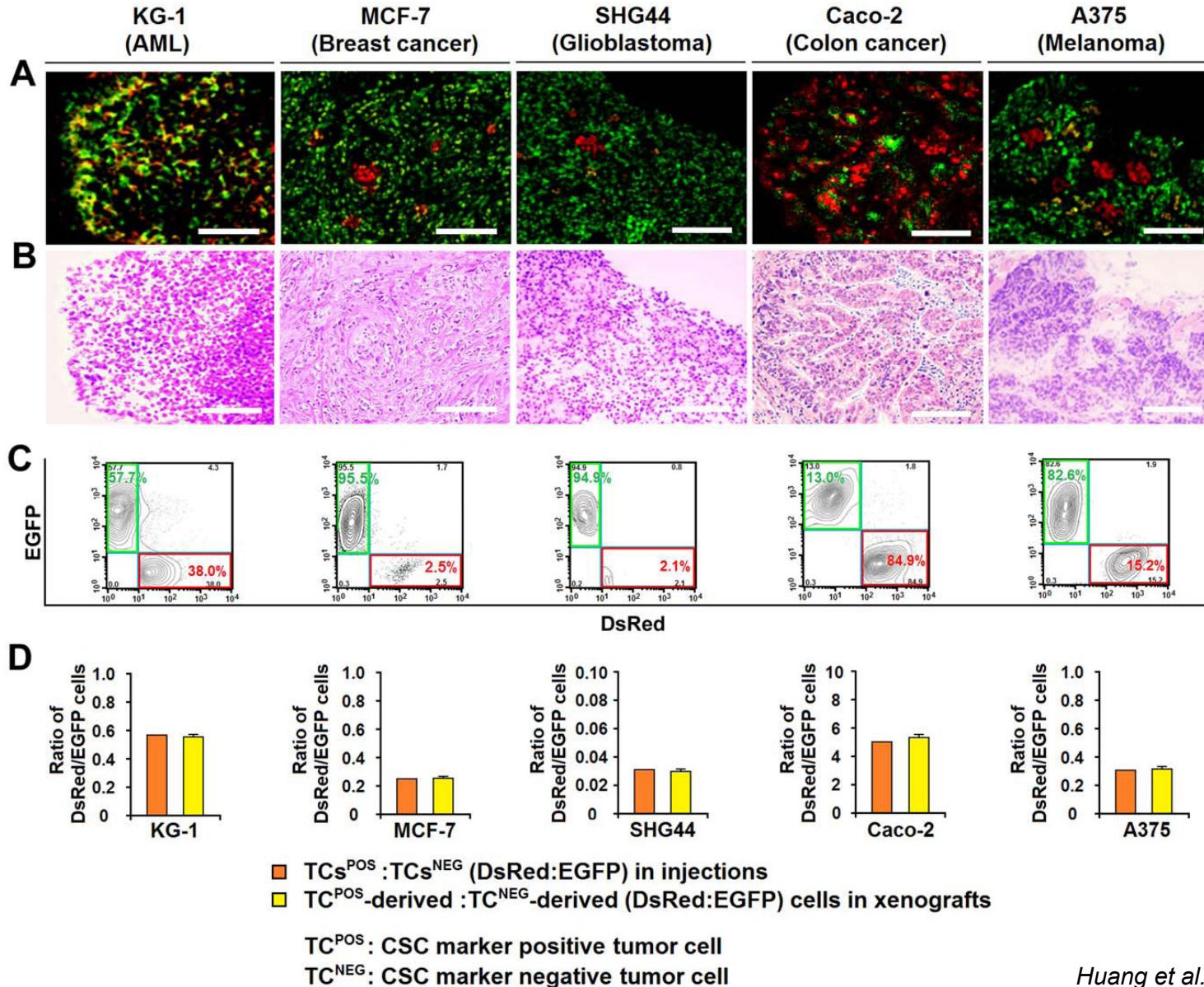
Elsa Quintana<sup>1</sup>\*, Mark Shackleton<sup>1</sup>\*, Michael S. Sabel<sup>2</sup>, Douglas R. Fullen<sup>3</sup>, Timothy M. Johnson<sup>4</sup> & Sean J. Morrison<sup>1</sup>

# Proliferation is similar in CSCs and nonCSCs but tumor induction upon transplantation differs



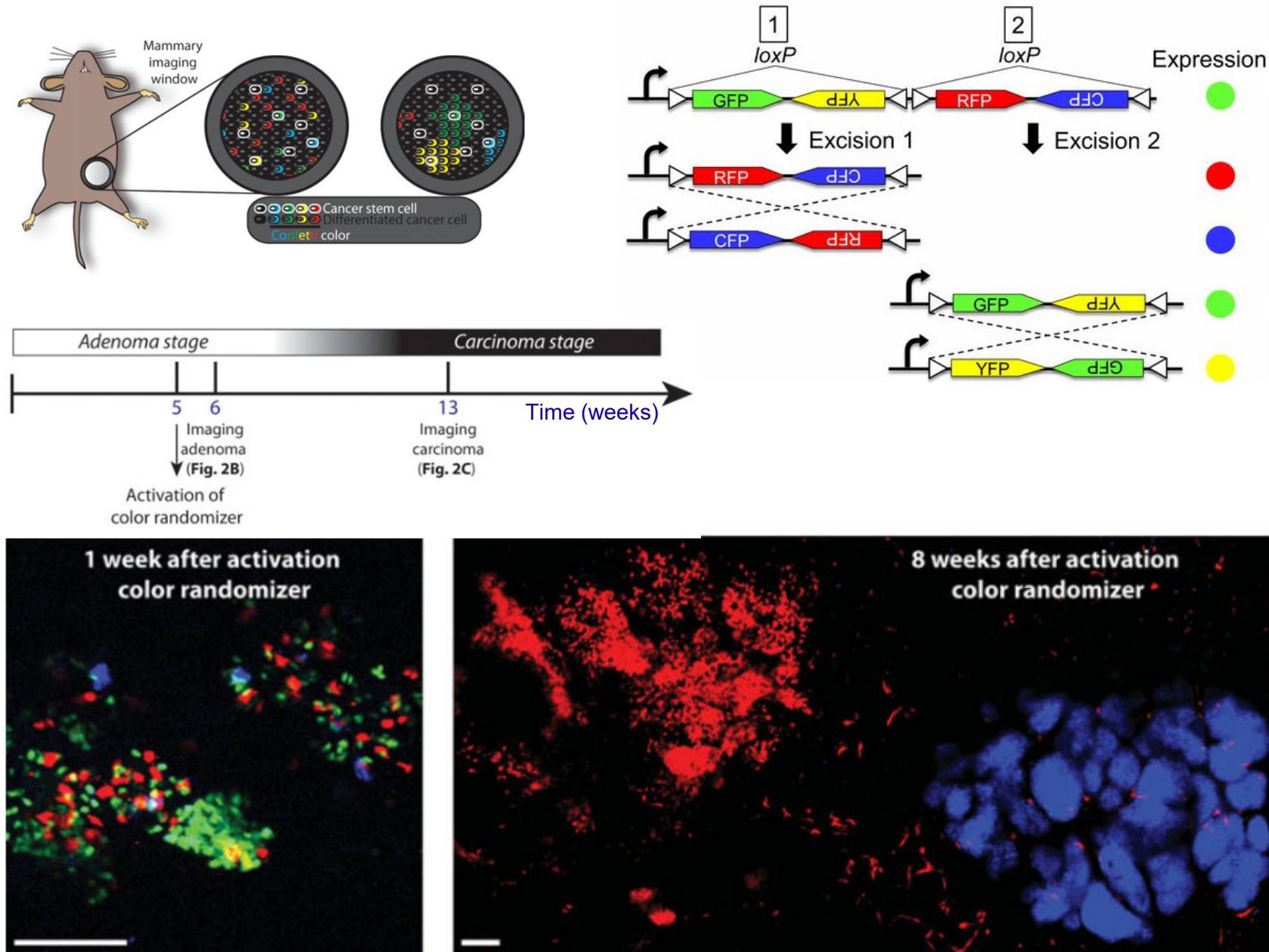
Cell line	KG-1		THP-1		HL60		K562	
Marker subset	CSC+	CSC-	CSC+	CSC-	CSC+	CSC-	CSC+	CSC-
Frequency of tumorigenic cells	1/2164	1/68078**	1/2164	1/280030**	1/4170	1/68078**	1/10720	1/135931**
Cell line	MCF-7		MDA-MB-231		SHG44		U251	
Marker subset	CSC+	CSC-	CSC+	CSC-	CSC+	CSC-	CSC+	CSC-
Frequency of tumorigenic cells	1/2164	1/280030**	1/4170	1/108957**	1/2164	1/135931**	1/2164	1/280030**
Cell line	HT-29		SW480		SW620		A375	
Marker subset	CSC+	CSC-	CSC+	CSC-	CSC+	CSC-	CSC+	CSC-
Frequency of tumorigenic cells	1/4170	1/280030**	1/2164	1/108957**	1/2164	1/280030**	1/4170	1/43259*

# Co-transplantation of CSCs and non CSCs allows nonCSCs to participate in cancer growth

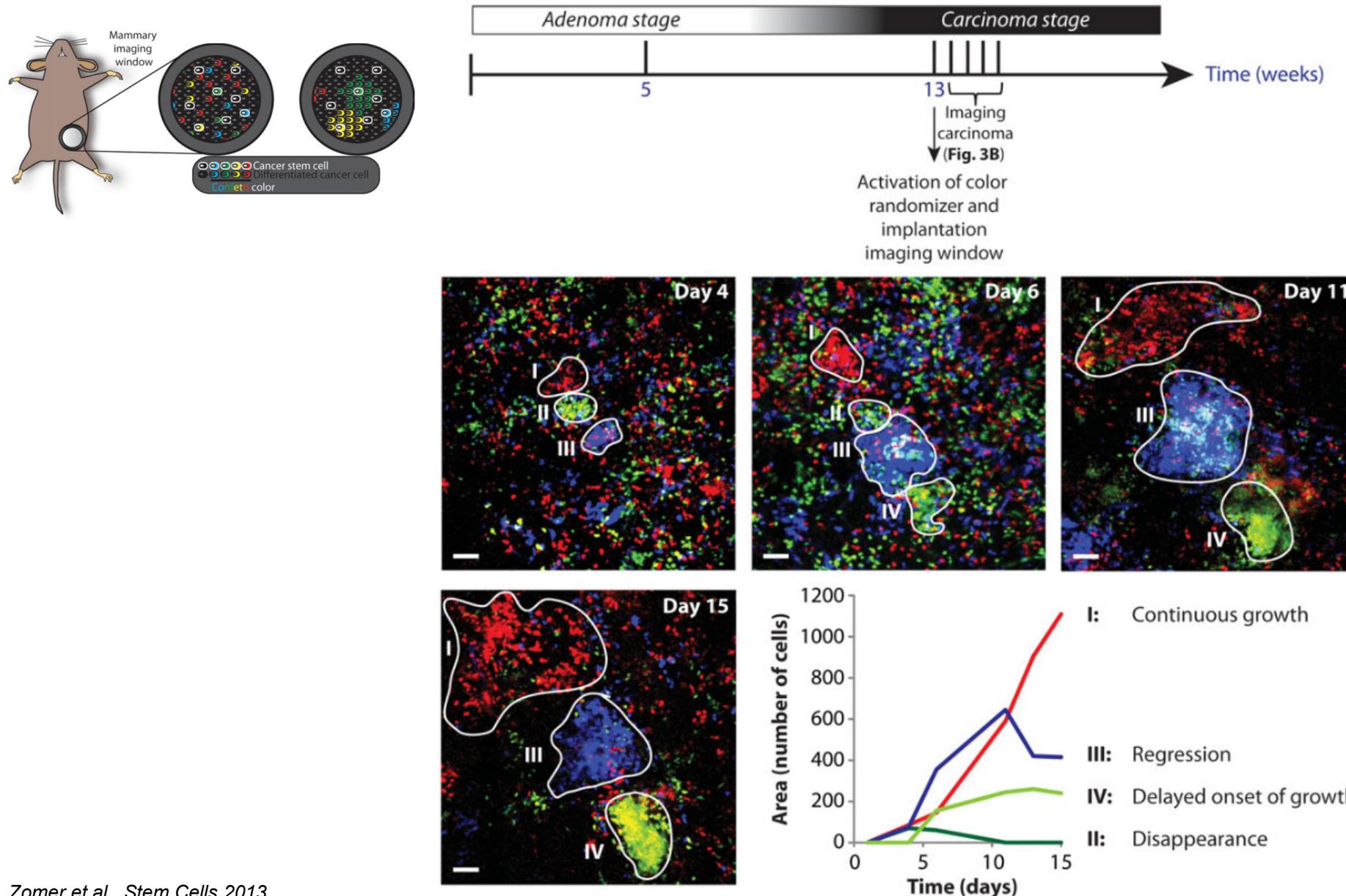


## **In vivo evidence for CSCs**

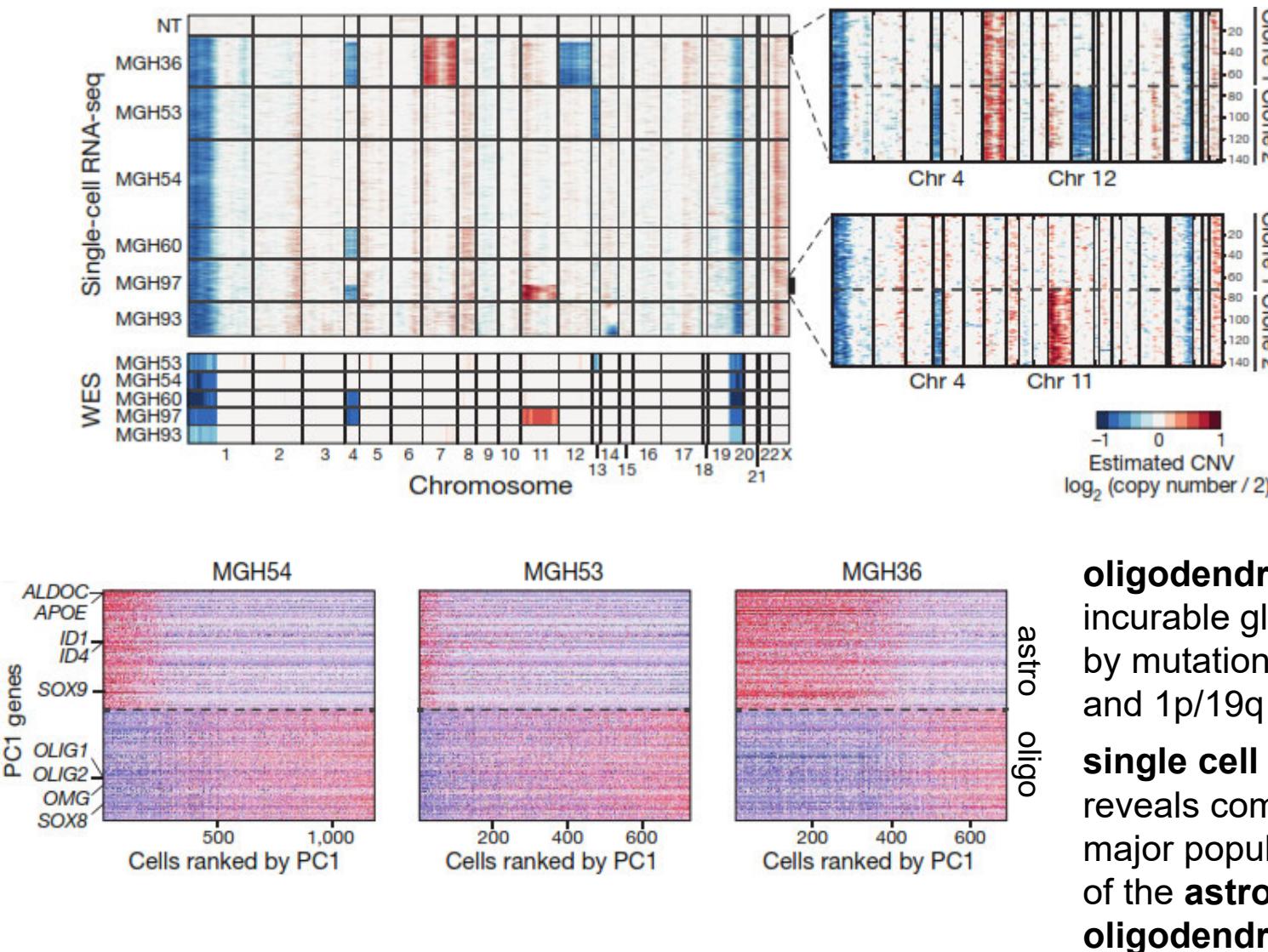
# Random labelling reveals CSC plasticity *in vivo*



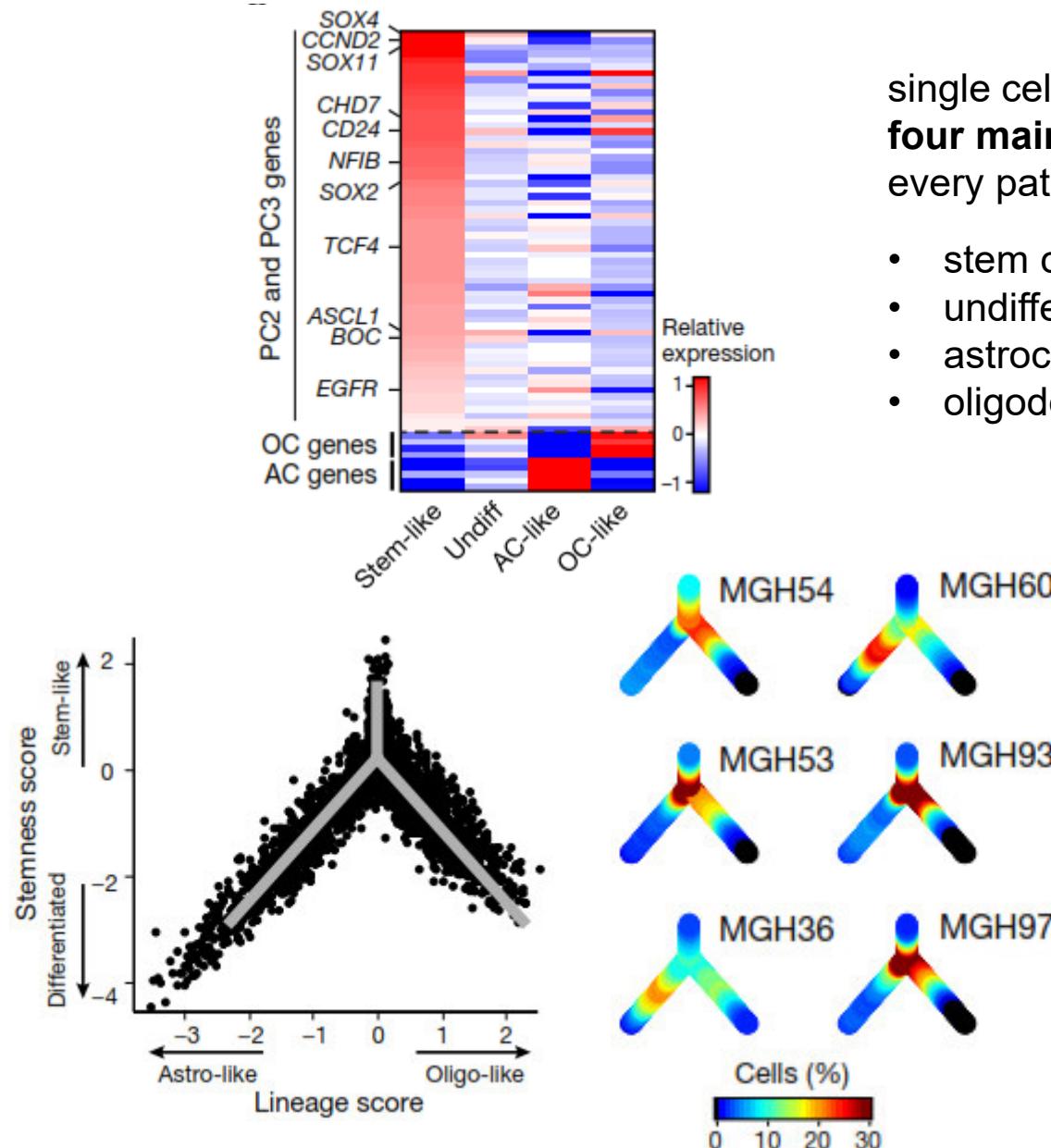
# Random labelling reveals CSC plasticity *in vivo*



# Direct Evidence for Cancer Hierarchy in Patient Samples



# Direct Evidence for Cancer Hierarchy in Patient Samples



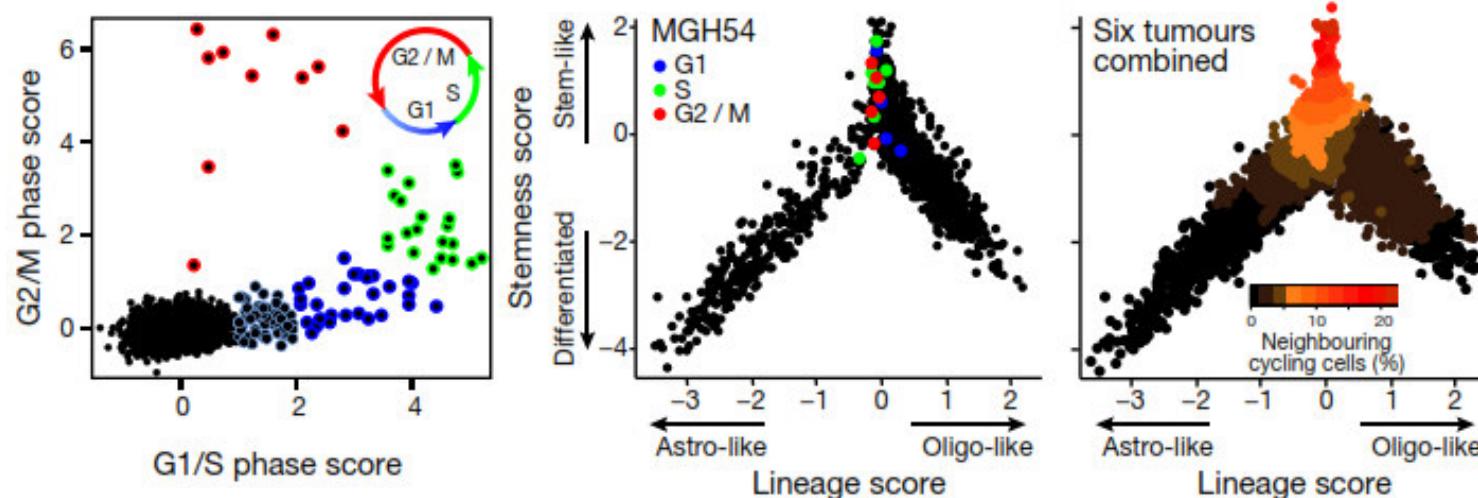
single cell analysis reveals  
**four main cell types** in  
every patient tumor:

- stem cell-like
- undifferentiated progenitors
- astrocyte-like
- oligodendrocyte-like

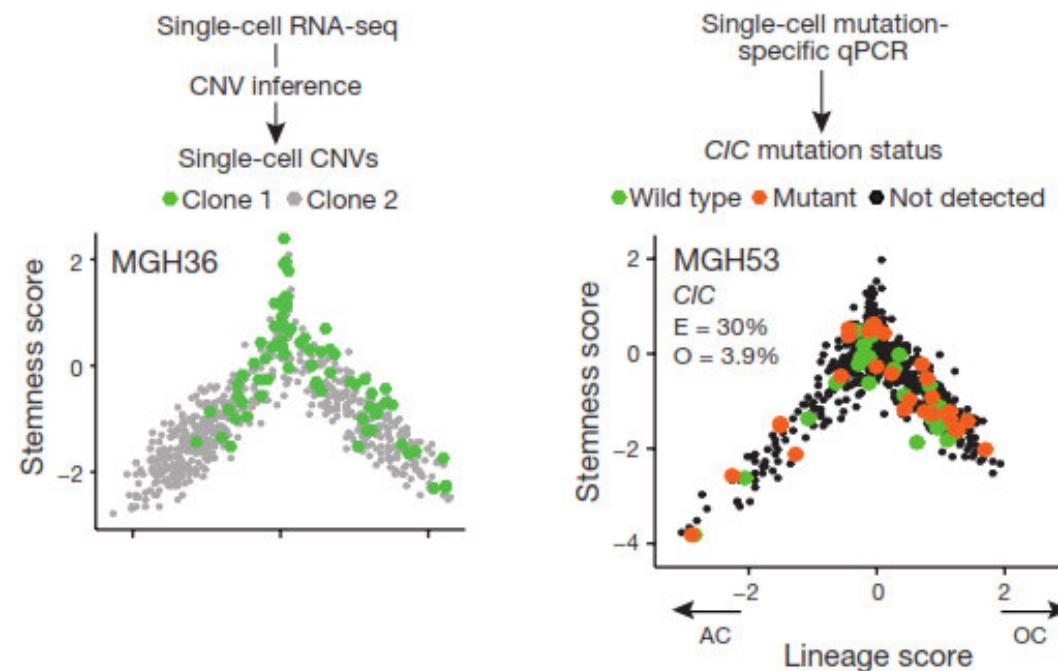
individual patients differ in  
the relative frequency of  
these four different cell  
types without loosing the  
general architecture

this architecture closely  
resembles the structure of  
the normal tissue

# Direct Evidence for Cancer Hierarchy in Patient Samples

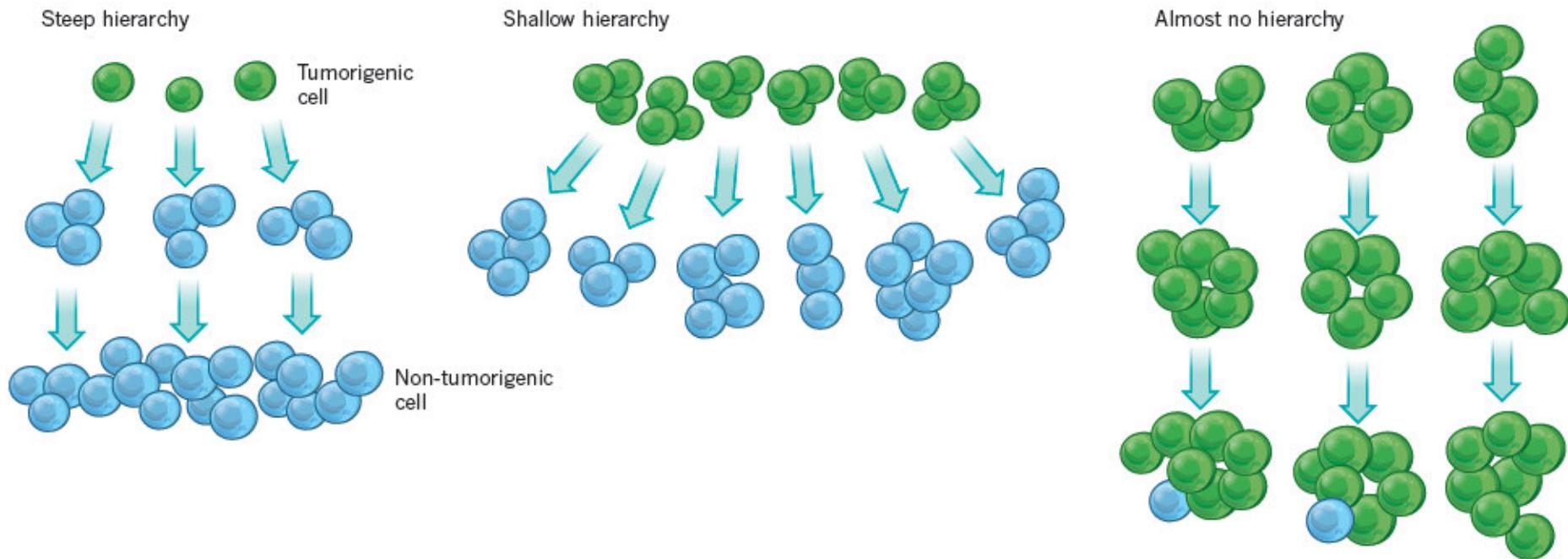


cell cycle activity  
is restricted to  
the stem and  
progenitor level



within the same patient genetically  
separate subclones co-exist  
which independently maintain the  
overall tissue architecture

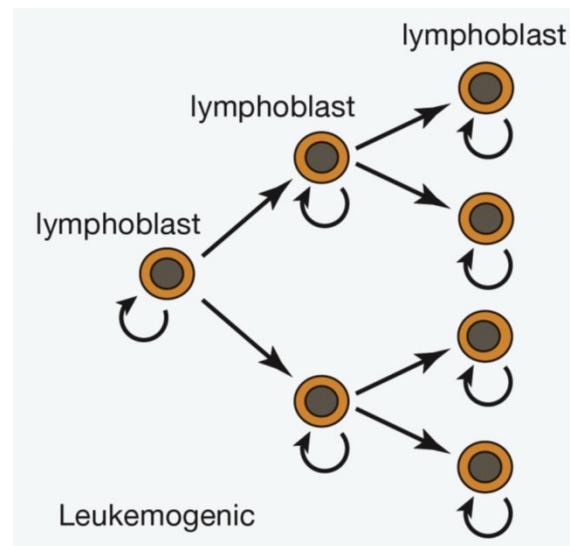
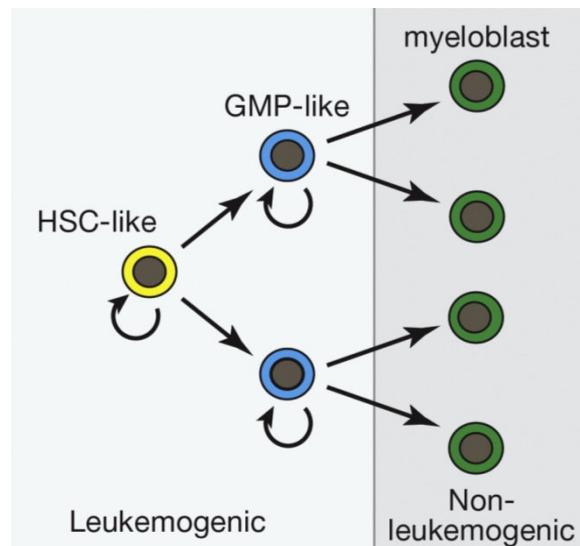
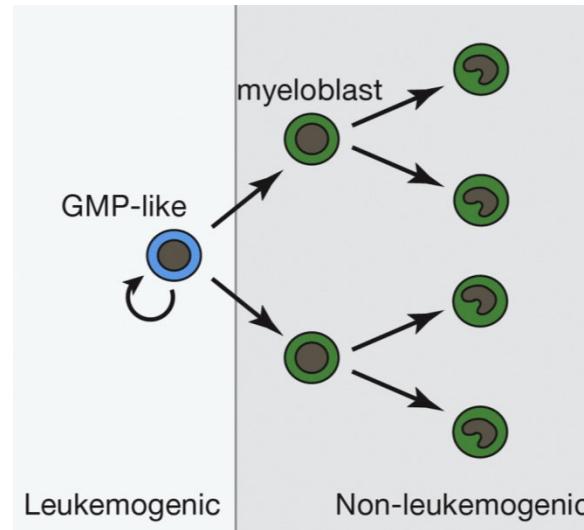
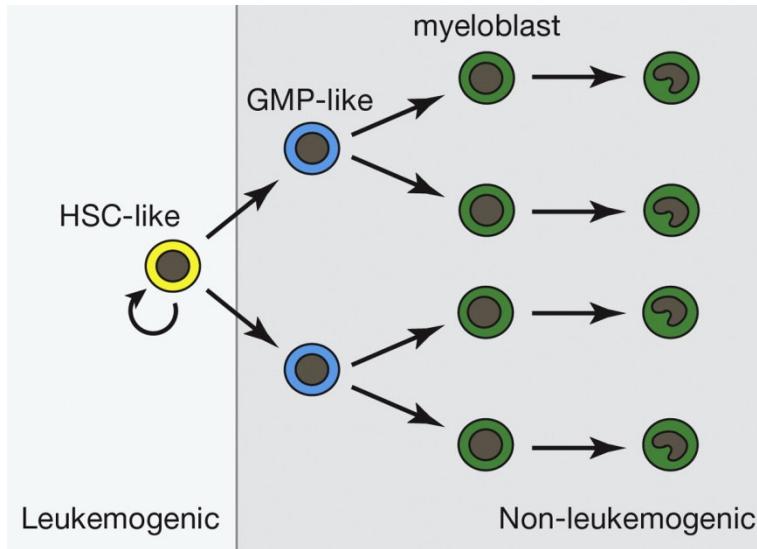
## Different levels of hierarchical organization



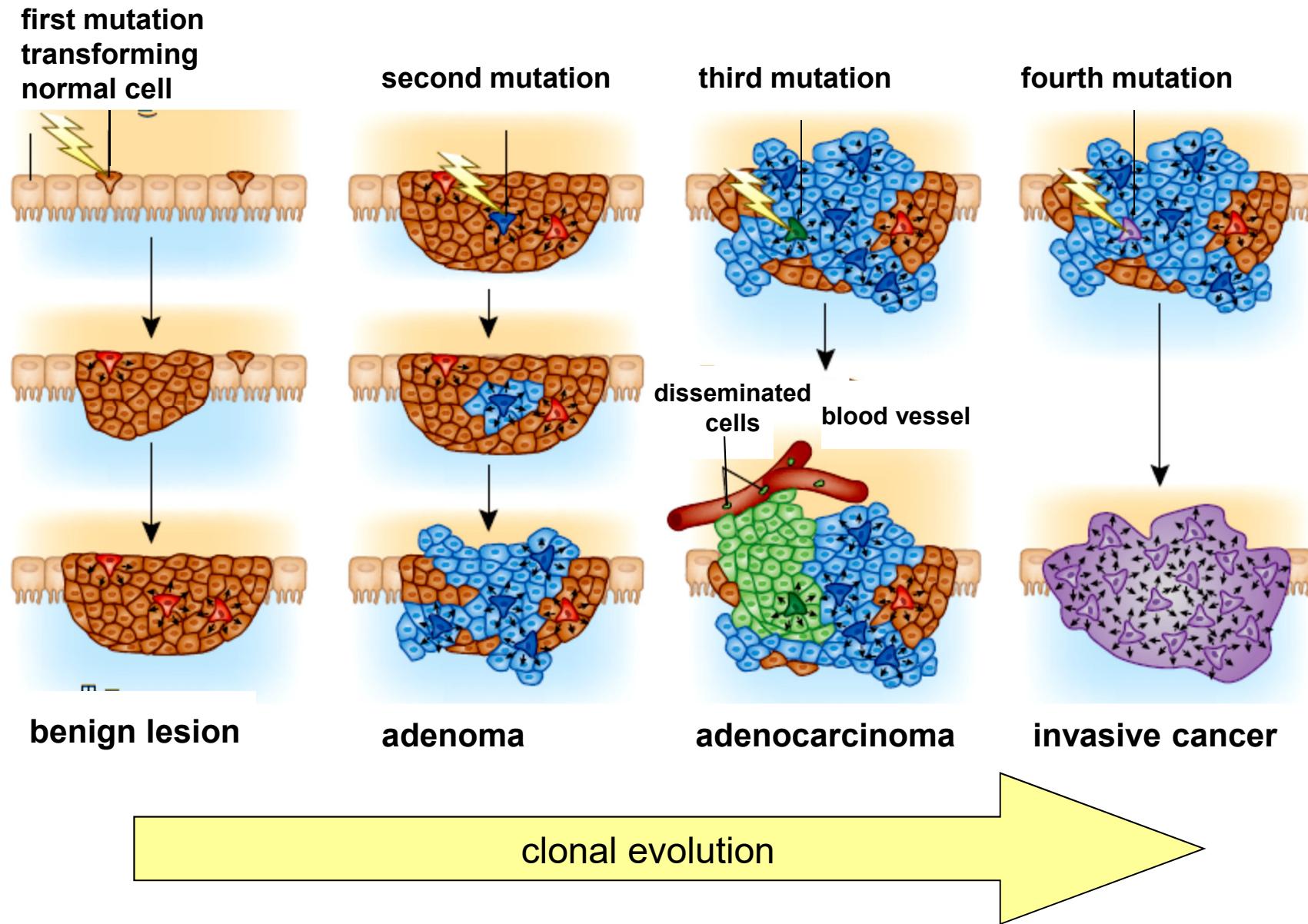
- Some hierarchies might be steep in which tumor-initiating cells are rare but give rise to numerous non-tumorigenic cells
- Other hierarchies might be shallow in which tumor-initiating cells are common but give rise to a small number of non-tumorigenic cells
- Some cancers may have almost no hierarchy, with very few non-tumorigenic cells.

The shallower the hierarchy, the lower the value of distinguishing between tumorigenic and non-tumorigenic cells in order to understand cancer biology and improve therapy.

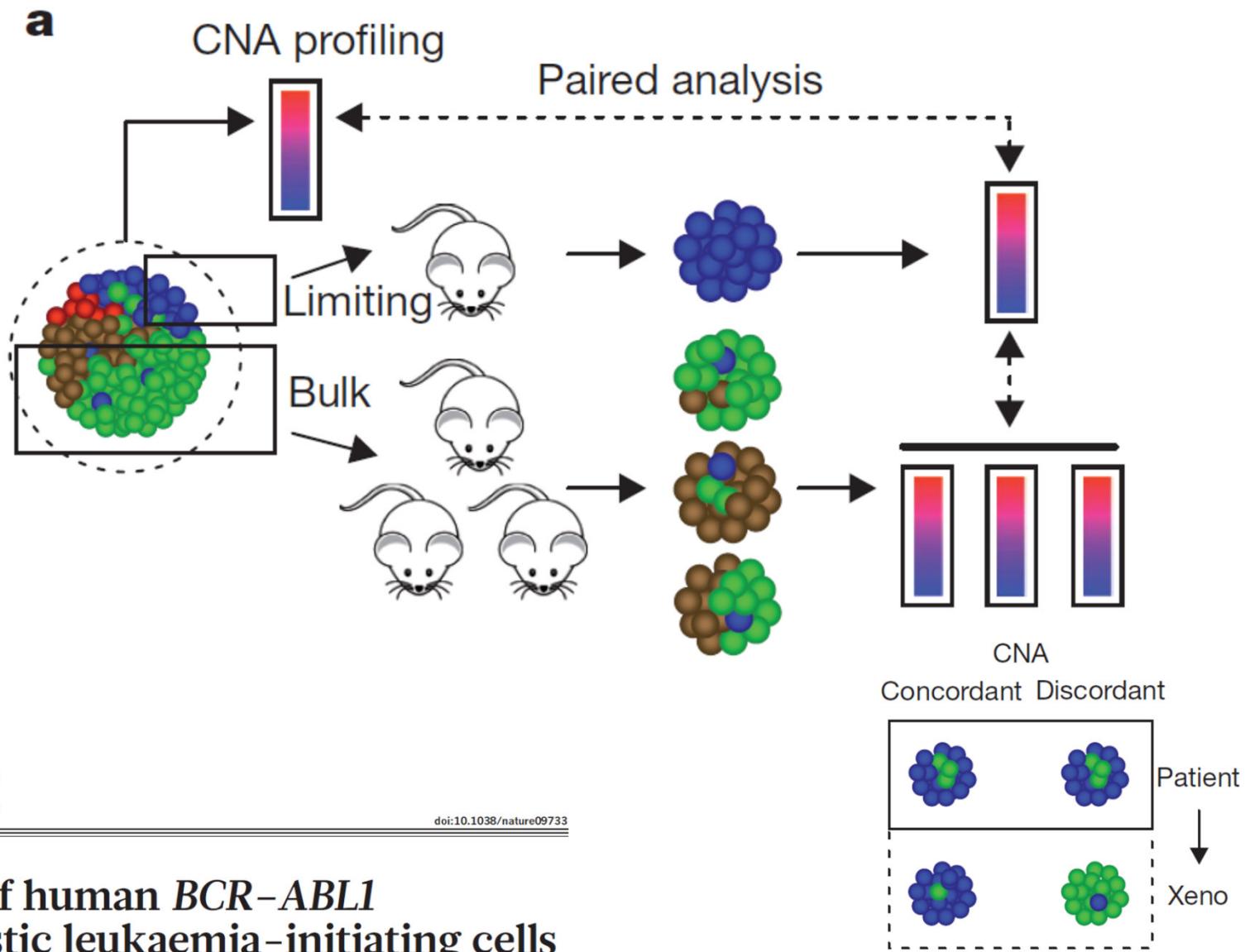
# Hierarchical Organization in Cancer



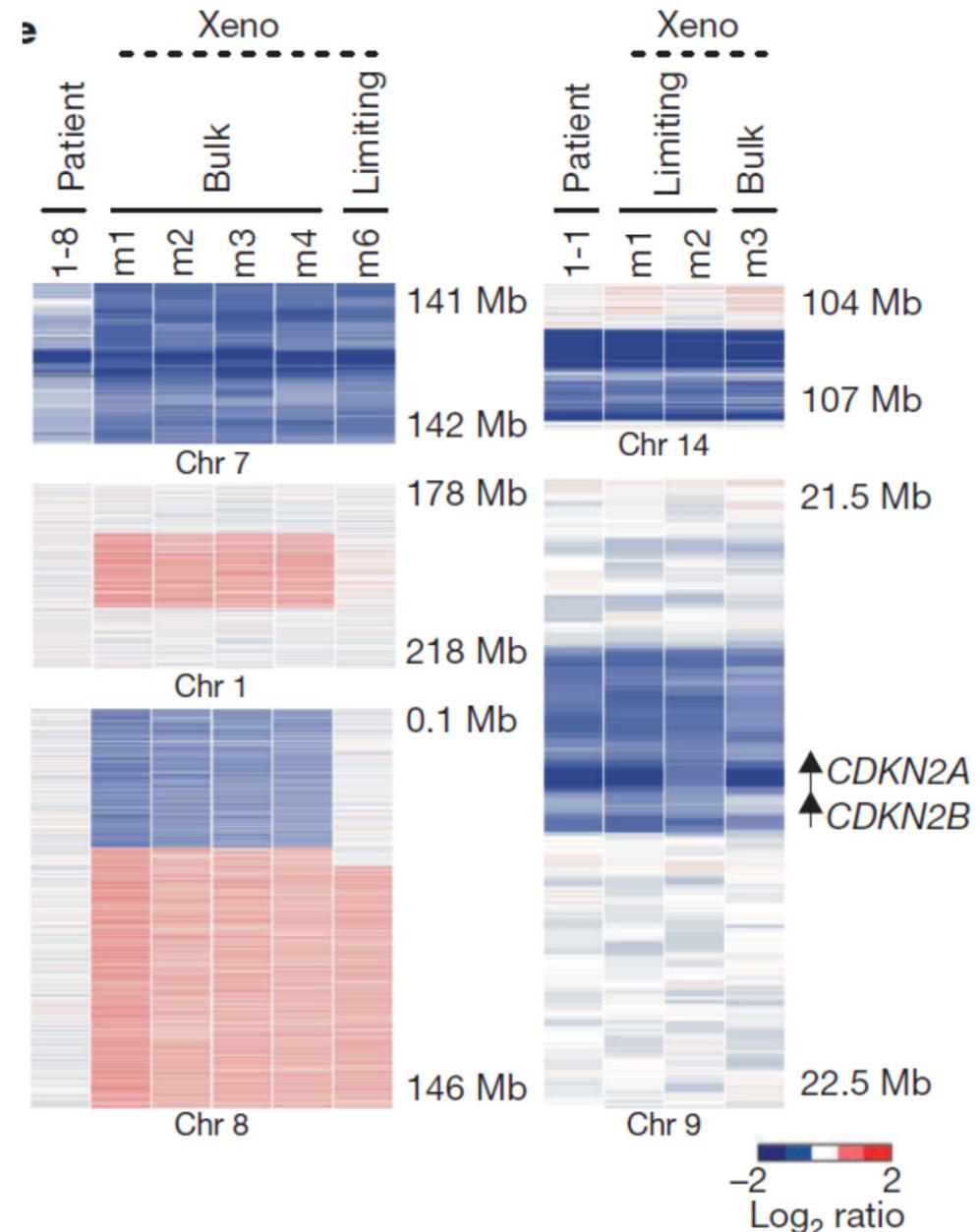
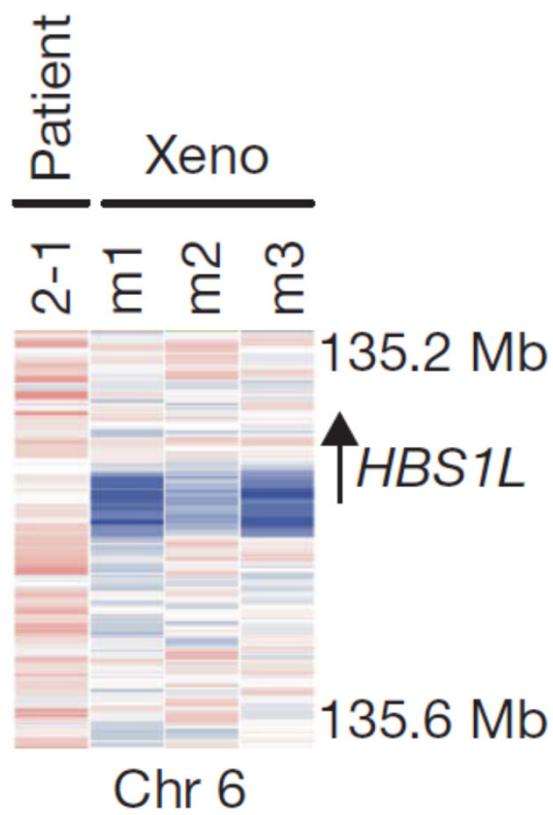
# Clonal evolution of Cancer



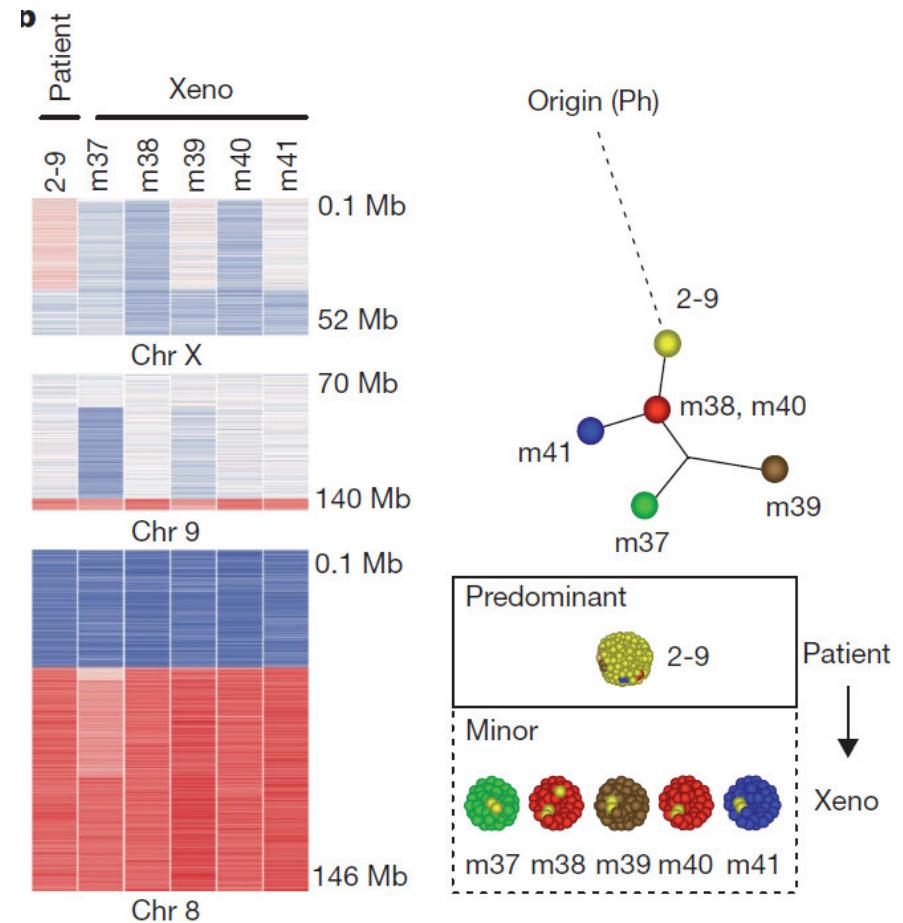
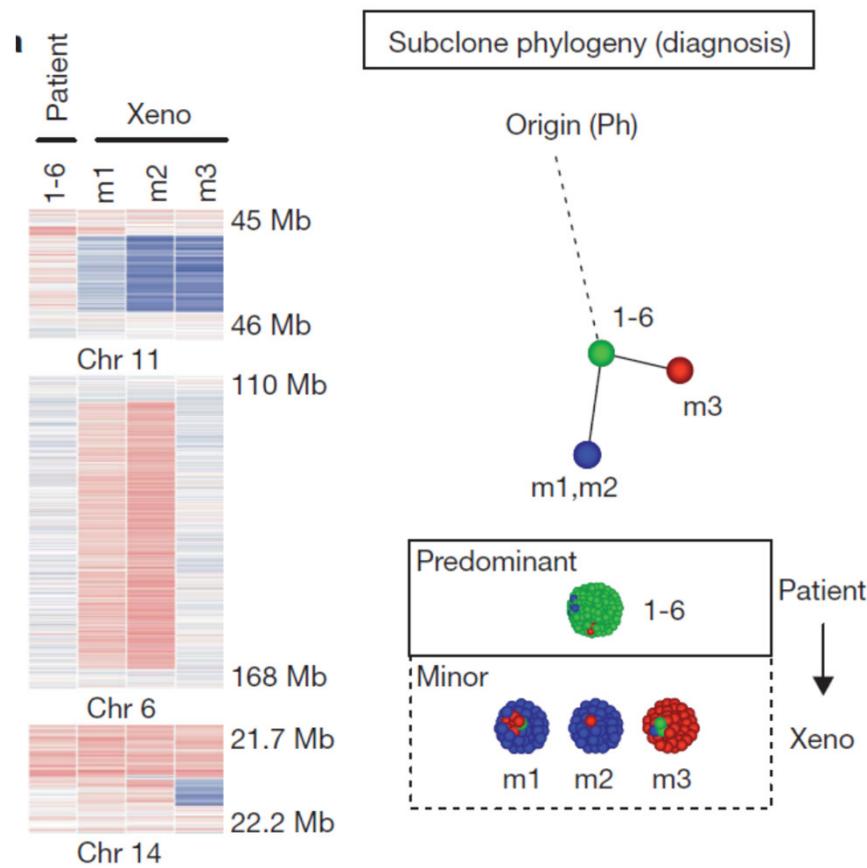
# Clonal evolution of CSCs



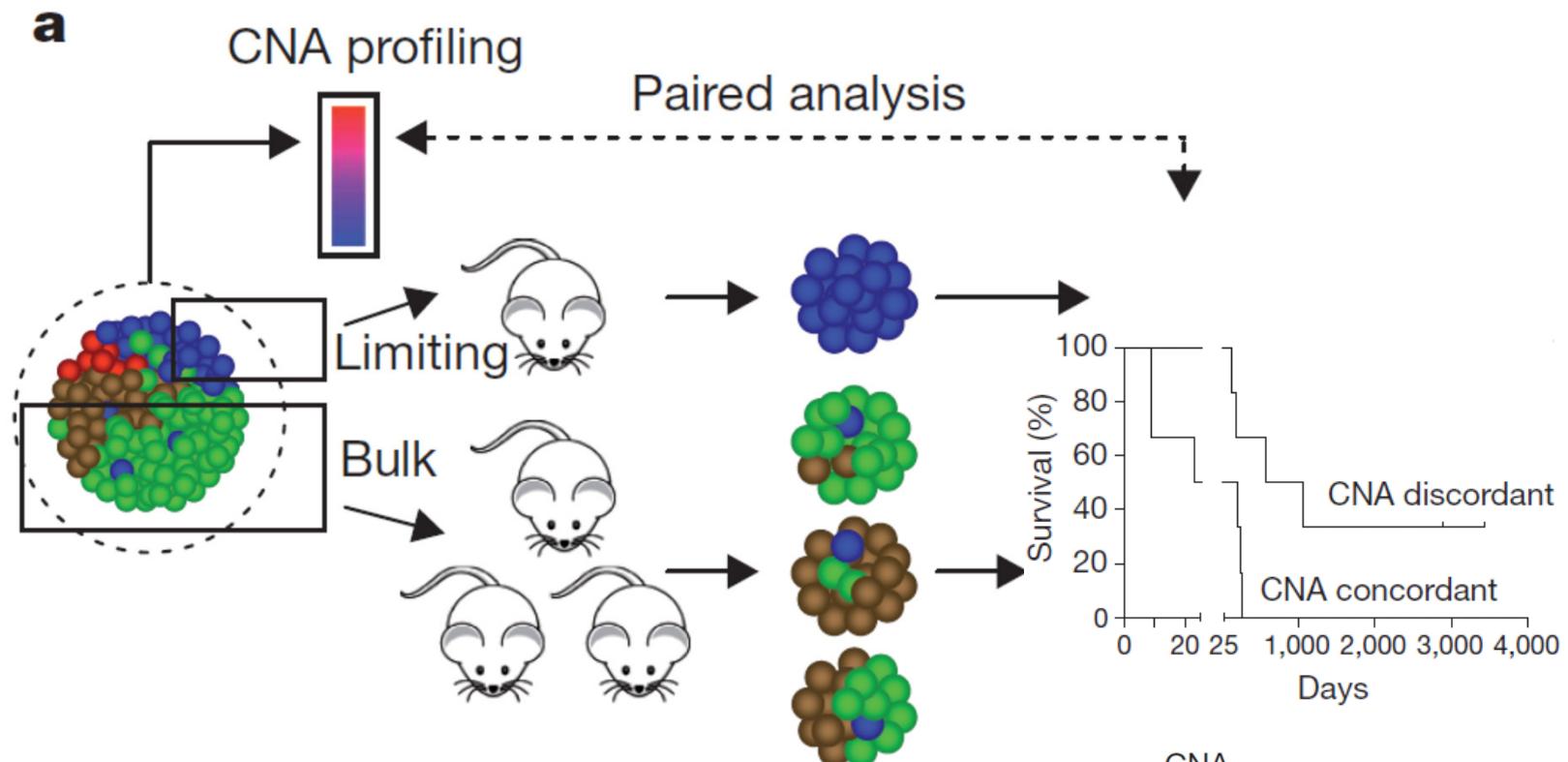
## Clonal evolution of CSCs: CML



# Clonal evolution of CSCs: CML



# Clonal evolution of CSCs



## ARTICLE

doi:10.1038/nature09733

### Evolution of human *BCR-ABL1* lymphoblastic leukaemia-initiating cells

Faiyaz Notta<sup>1,2\*</sup>, Charles G. Mullighan<sup>3\*</sup>, Jean C. Y. Wang<sup>1,4</sup>, Armando Poepl<sup>1</sup>, Sergei Doulatov<sup>1,2</sup>, Letha A. Phillips<sup>3</sup>, Jing Ma<sup>5</sup>, Mark D. Minden<sup>4</sup>, James R. Downing<sup>3</sup> & John E. Dick<sup>1,2</sup>

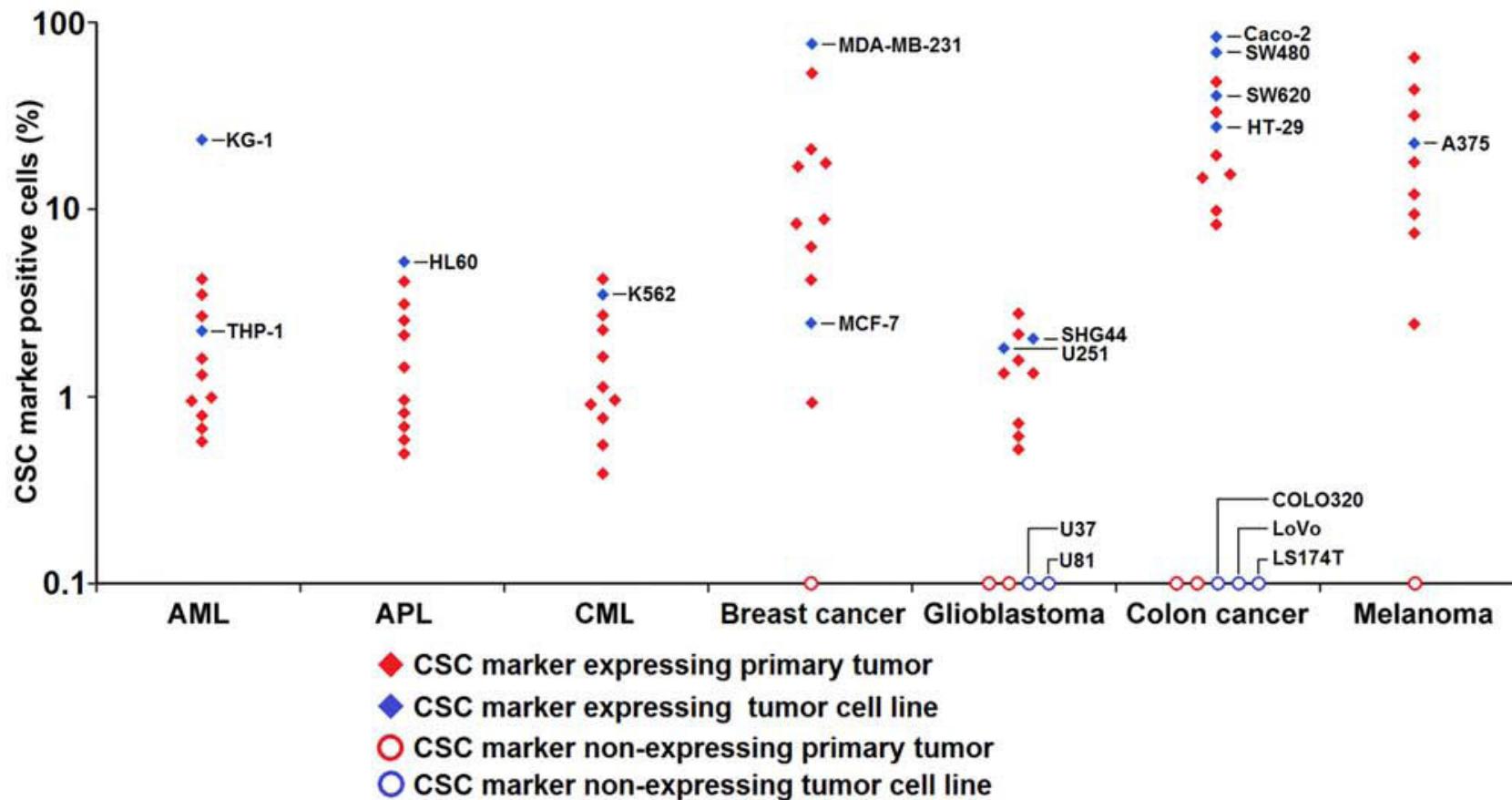
## **Markers to identify CSCs**

# Markers to isolate CSCs

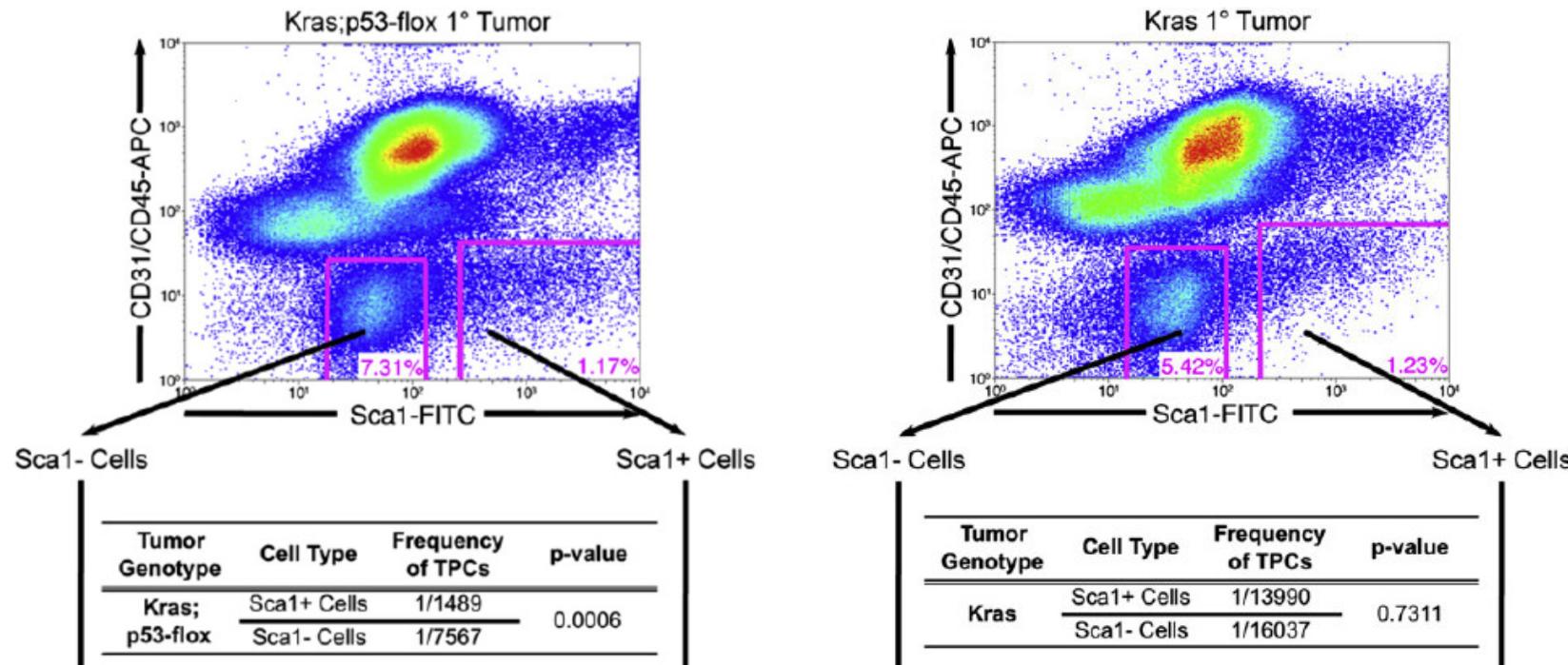
Breast	Colon	Glioma	Liver	Lung	Melanoma	Ovarian	Pancreatic	Prostate
ALDH1	ABCB5	CD15	CD13	ABCG2	ABCB5	CD24	ABCG2	ALDH1
CD24	ALDH1	CD90	CD24	ALDH1	ALDH1	CD44	ALDH1	CD44
CD44	$\beta$ -catenin	CD133	CD44	CD90	CD20	CD117	CD24	CD133
CD90	activity	$\alpha$ 6-integrin	CD90	CD117	CD133	CD133	CD44	CD166
CD133	CD24	nestin	CD133	CD133	CD271		CD133	$\alpha$ 2 $\beta$ 1-integrin
Hedgehog-Gli activity	CD26						c-Met	$\alpha$ 6-integrin
$\alpha$ 6-integrin	CD29						CXCR4	Trop2
	CD44						Nestin	
	CD133						Nodal-Activin	
	CD166							
	LGR5							

- all current markers are not specific for CSCs, but are expressed by many other cells outside of the tumor
- this makes direct targeting of the CSC population using these markers unlikely
- most current markers are not stem cell markers for the normal tissue stem cells where the tumor derives from
  - exceptions: CD34/CD38 for human leukemia/HSC  
CD133 for human glioblastoma/neuronal stem cells  
Sca1/c-kit for mouse leukemia/HSC  
CD34 for mouse SCC/skin stem cells
- nearly all current markers play no functional role in CSCs or normal stem cells
  - exceptions: CD44 for glioblastoma and colon cancer  
Sca1/c-kit for mouse leukemia/HSC  
CD34 for mouse SCC
- markers that were originally characterized in a limited number of tumours have often been assumed to be generalizable. Such markers have frequently been used in other tumours, or even in cell lines, without independent confirmation that the markers were informative in these contexts
- in some cancers CSCs express different markers depending on the causative mutations

## CSC marker expression can be heterogeneous



# Within one tissue, CSC markers may depend on the genetic cause of tumorigenesis



Tumor Genotype	Cell Type	Tumor formation
EGFR	Sca1+ Cells	0/4
	Sca1- Cells	7/7

Cell Stem Cell  
Short Article



## Primary Tumor Genotype Is an Important Determinant in Identification of Lung Cancer Propagating Cells

Stephen J. Curtis,<sup>1,2,3</sup> Kerstin W. Sinkevicius,<sup>1,2,3</sup> Danan Li,<sup>4,5,6</sup> Allison N. Lau,<sup>1,2,3</sup> Rebecca R. Roach,<sup>1,2,3</sup> Raffaella Zamponi,<sup>1,2,3</sup> Amber E. Woolfenden,<sup>7</sup> David G. Kirsch,<sup>8</sup> Kwok-Kin Wong,<sup>4,5,6</sup> and Carla F. Kim<sup>1,2,3,\*</sup>

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<sup>2</sup>Department of Genetics, Harvard Medical School, Boston, MA 02115, USA

<sup>3</sup>Harvard Stem Cell Institute, Cambridge, MA 02138, USA

<sup>4</sup>Department of Medical Oncology, Dana-Farber Cancer Institute, 44 Binney Street, Boston, MA 02115, USA

<sup>5</sup>Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA 02115, USA

<sup>6</sup>Ludwig Center at Dana-Farber/Harvard Cancer Center, 44 Binney Street, Boston, MA 02115, USA

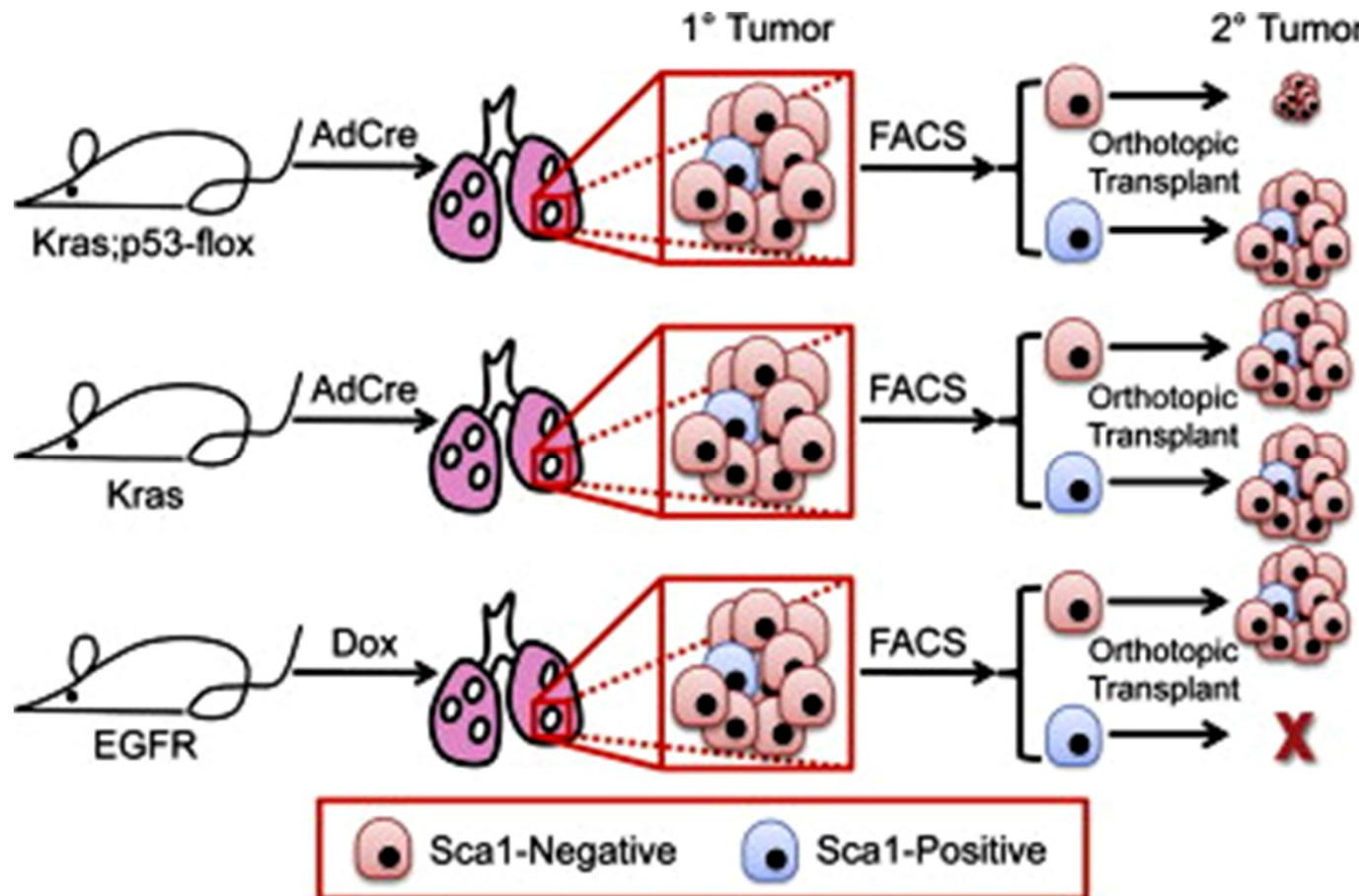
<sup>7</sup>Novartis Institutes for BioMedical Research, Inc., Cambridge, MA 02139

<sup>8</sup>Departments of Radiation Oncology and Pharmacology & Cancer Biology, Duke University Medical Center, Durham, NC 27708, USA

\*Correspondence: carla.kim@childrens.harvard.edu

DOI 10.1016/j.jstem.2010.05.021

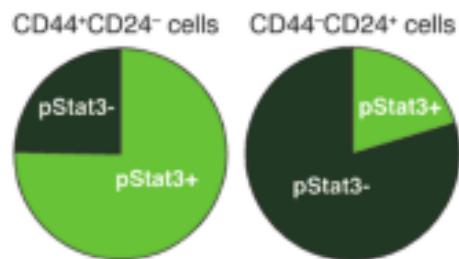
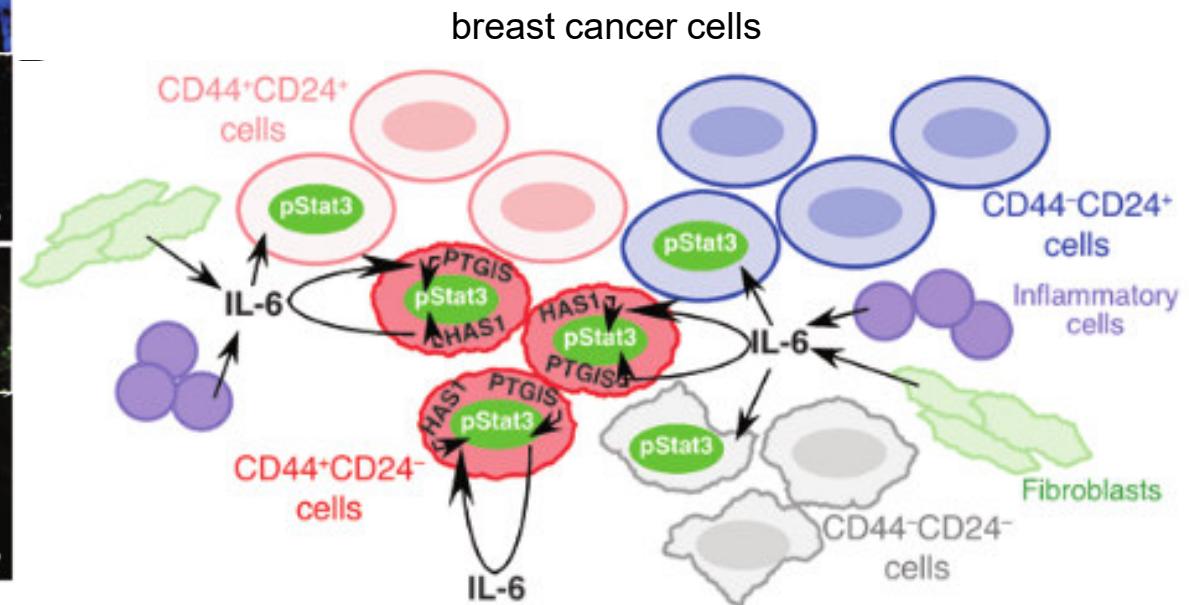
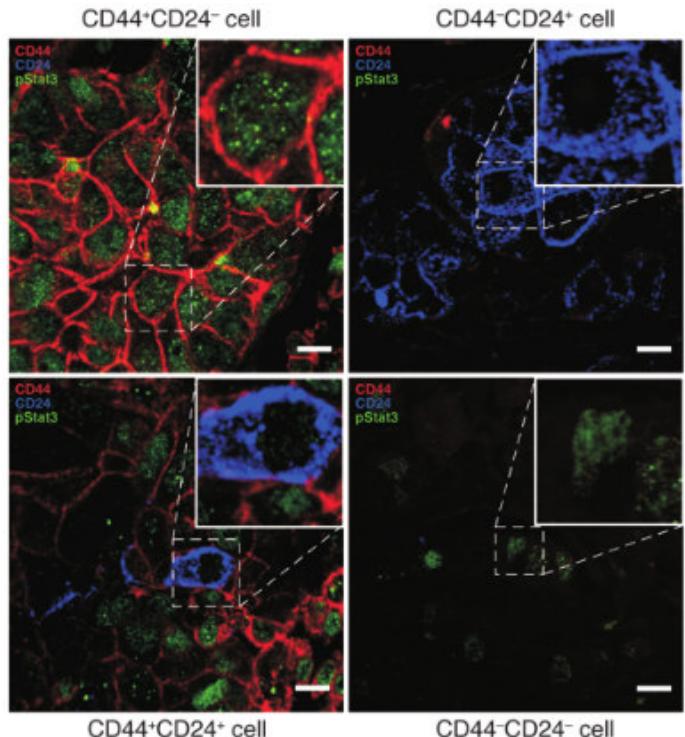
## Within one tissue, CSC markers may depend on the genetic cause of tumorigenesis



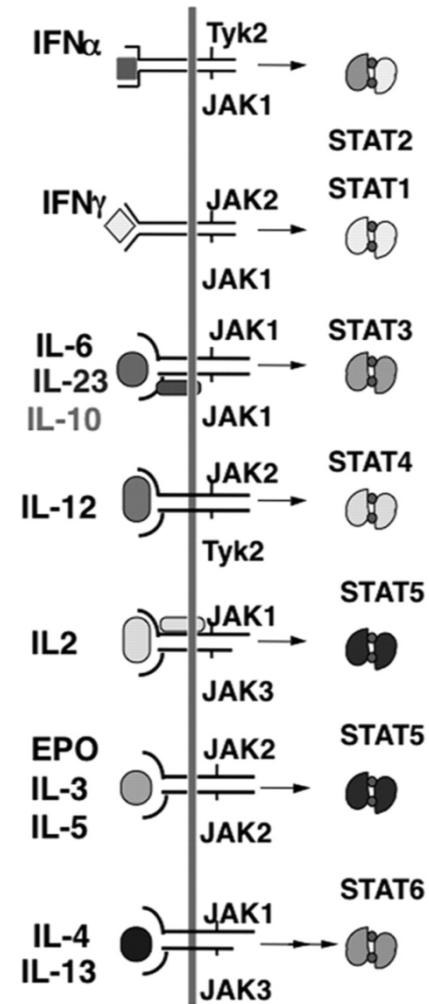
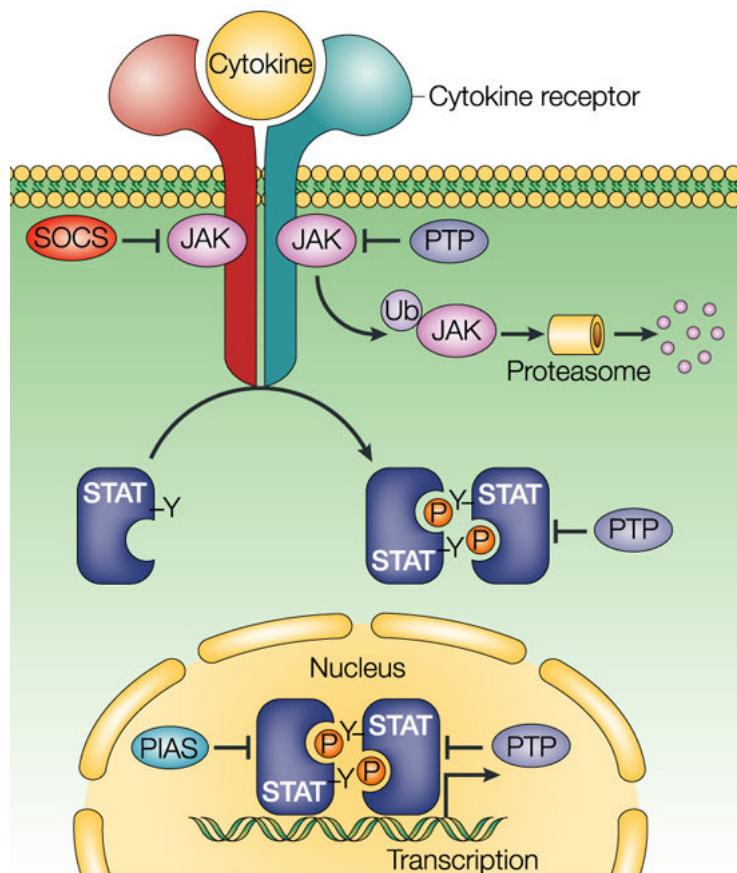
## **Stability of the CSC phenotype**

# Stability and Induction of CSCs

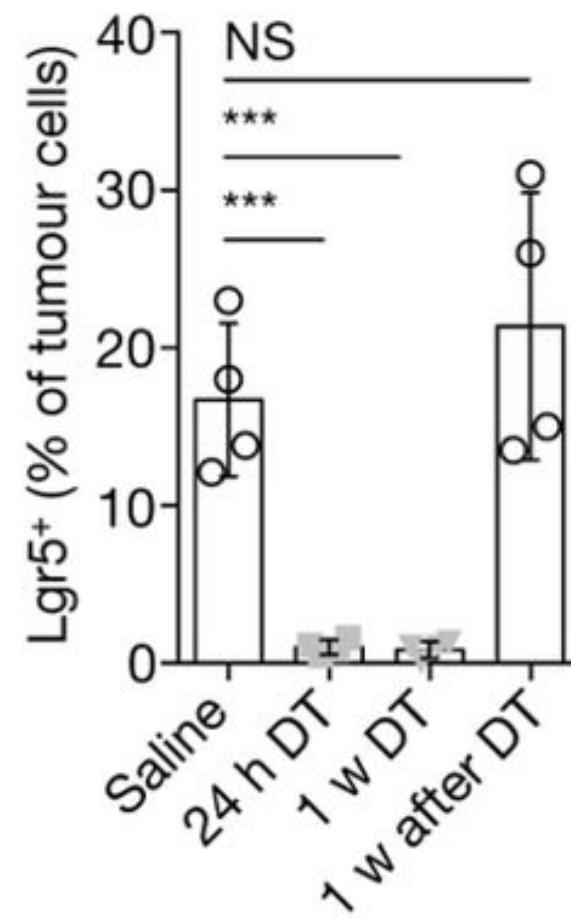
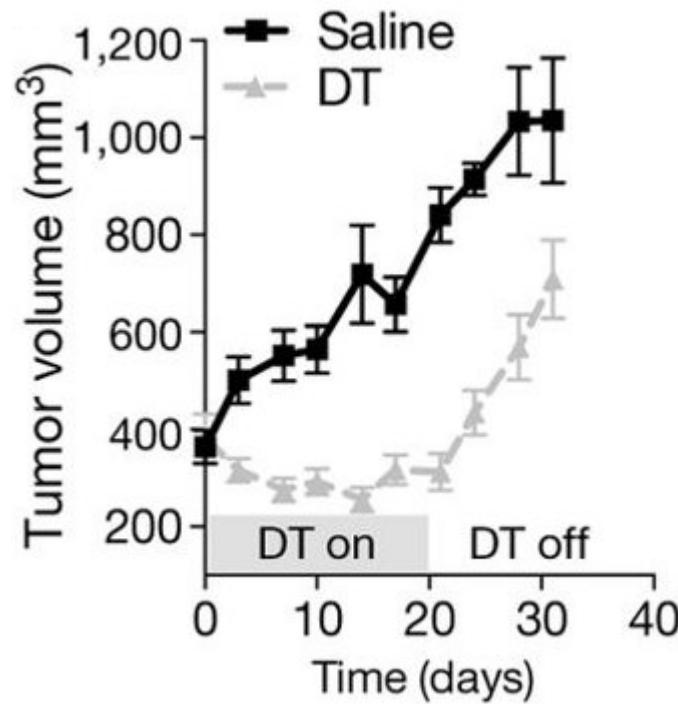
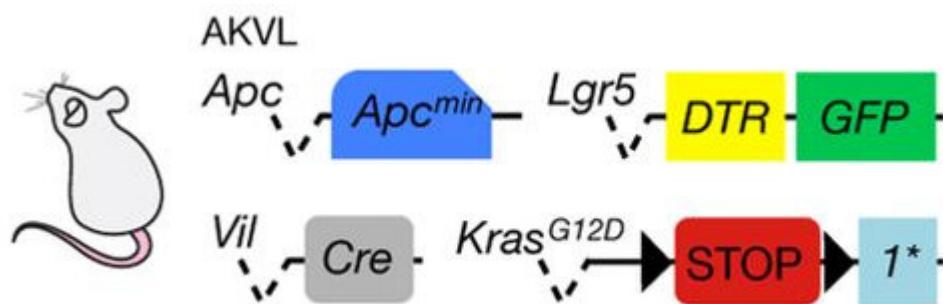
any experiment only represents a snapshot in time: can nonCSCs generate CSCs ?



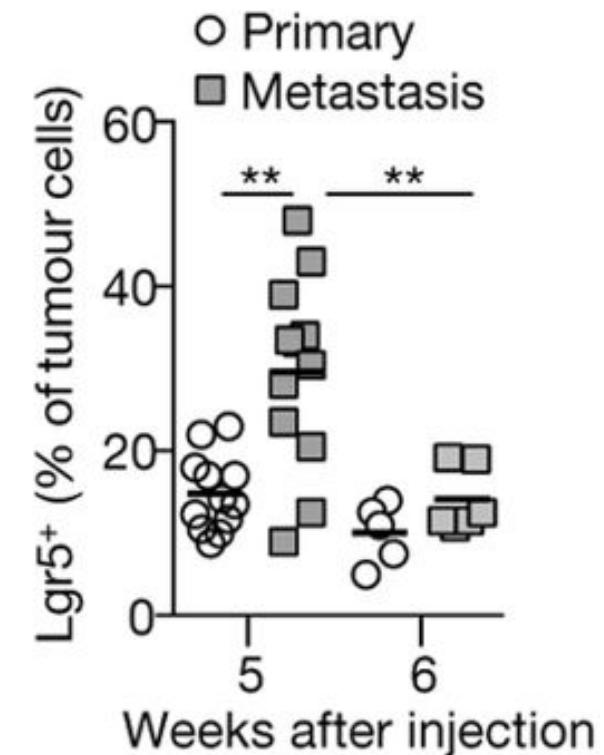
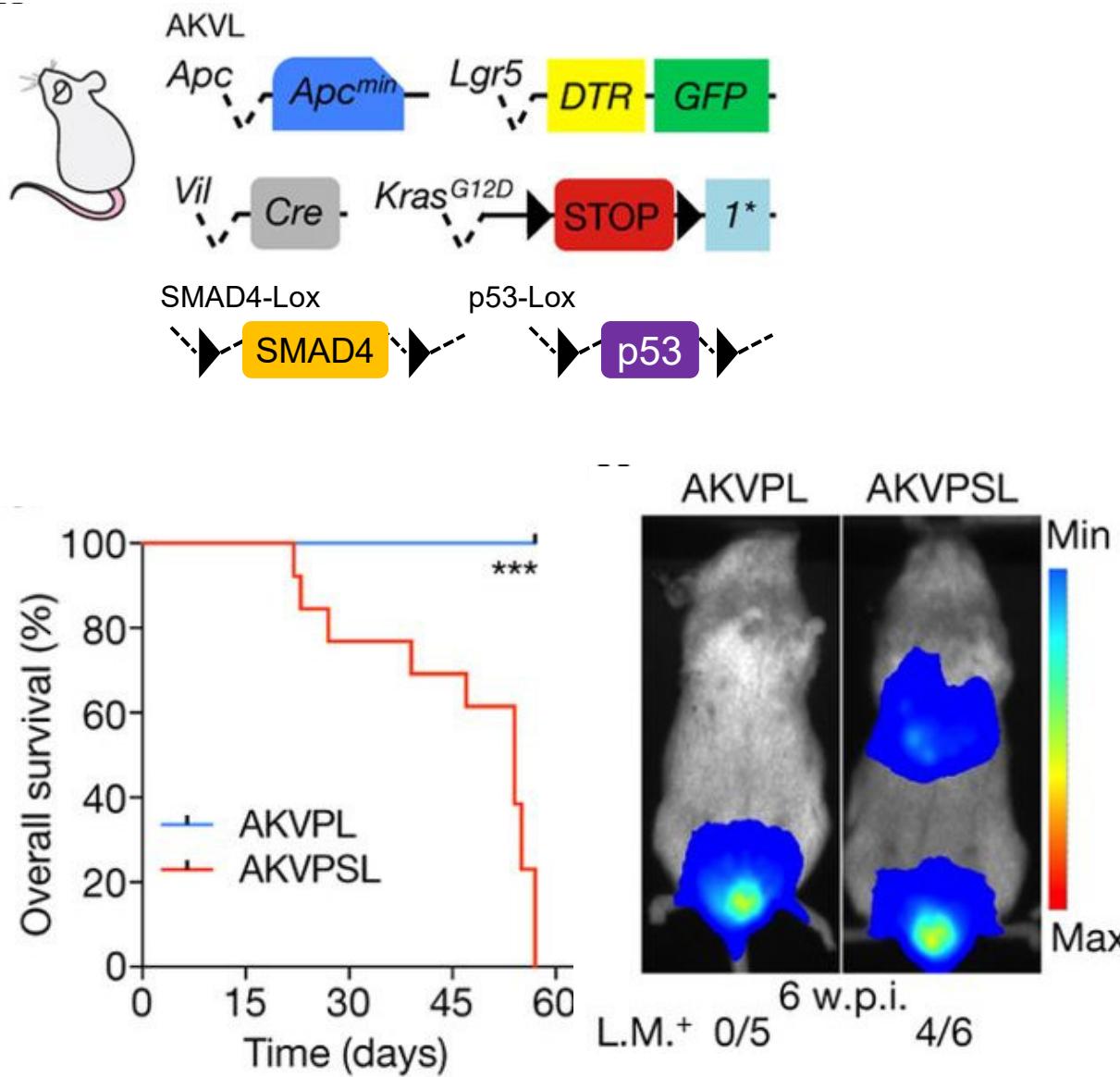
# Stability and Induction of CSCs



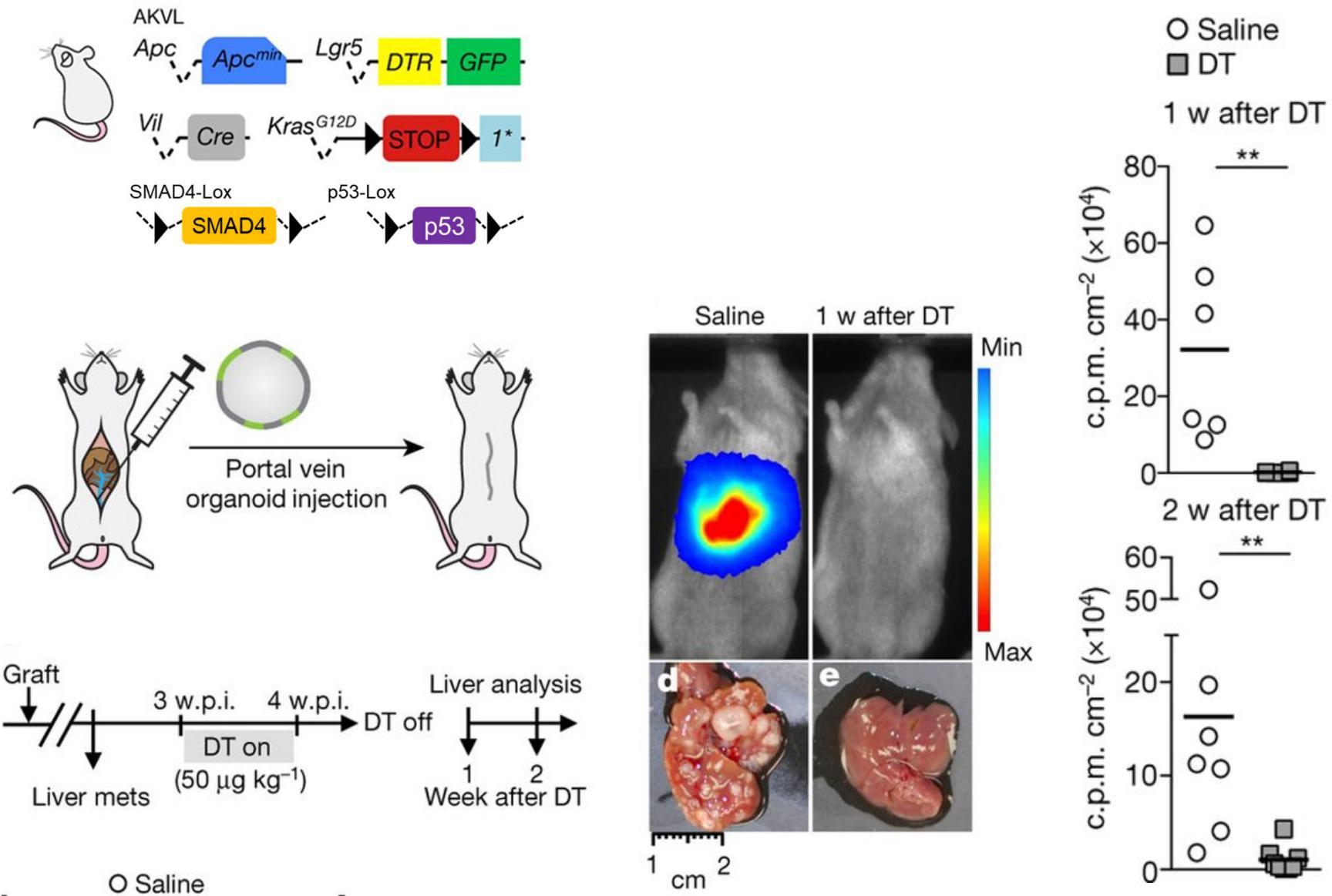
## *In vivo ablation of CSC is only transient*



# CSC frequency during metastasis formation



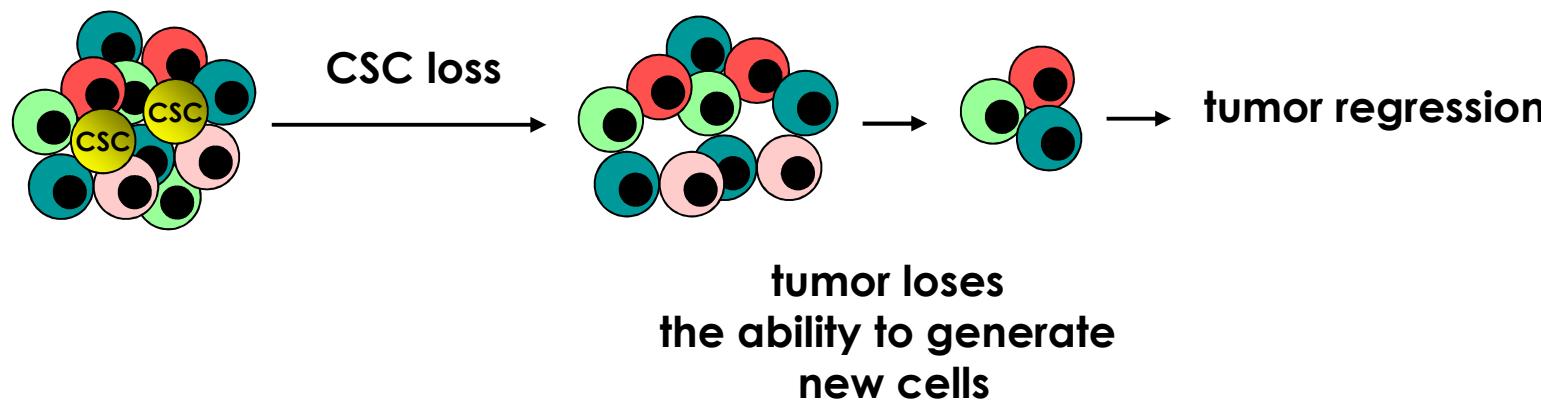
# *In vivo* ablation of CSC can prevent metastasis formation



# Targeting CSCs

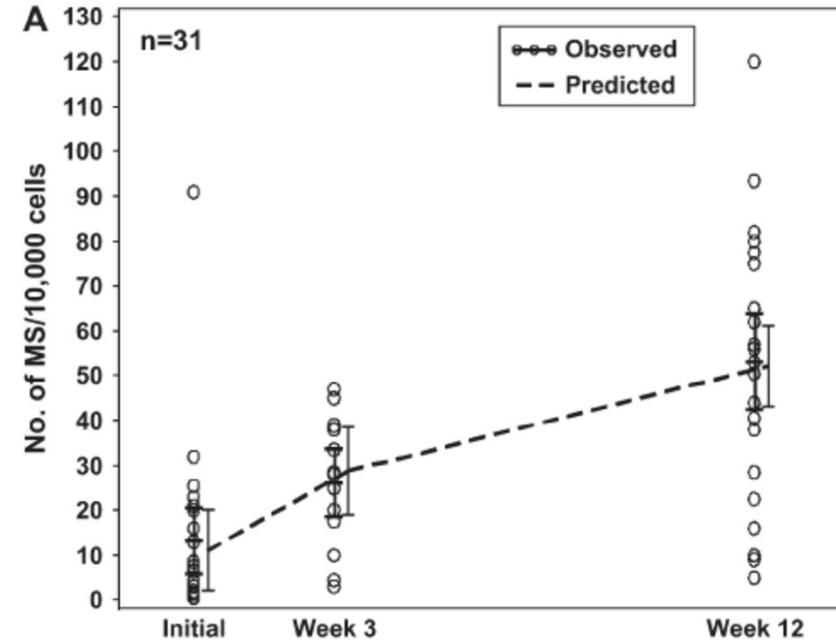
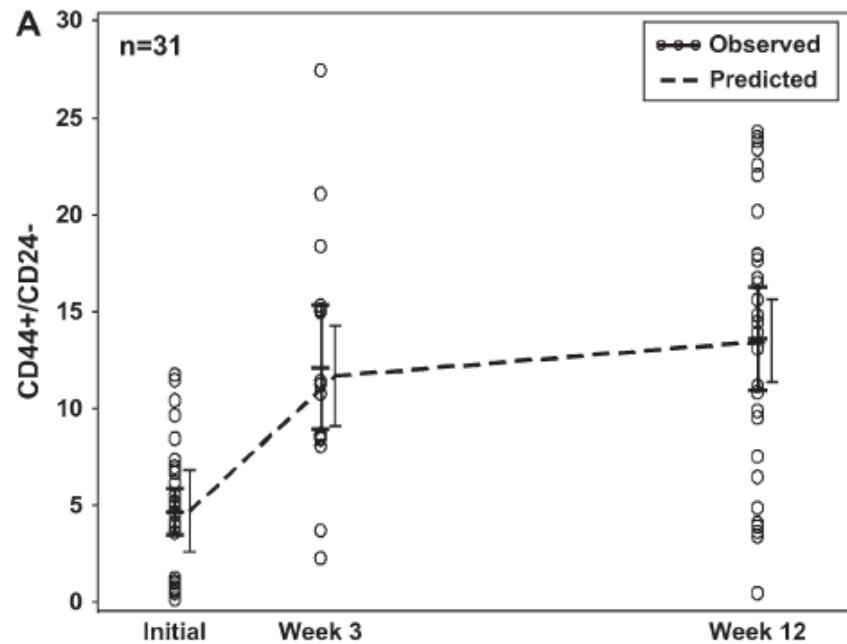
# Promises and challenges of the cancer stem cell concept

Targeted elimination of CSC will induce complete tumor regression and prevent recurrence



**It is central to identify signaling pathways which are critical for cancer stem cell maintenance, however are not essential for normal tissue homeostasis**

# Resistance to therapy breast cancer chemotherapy

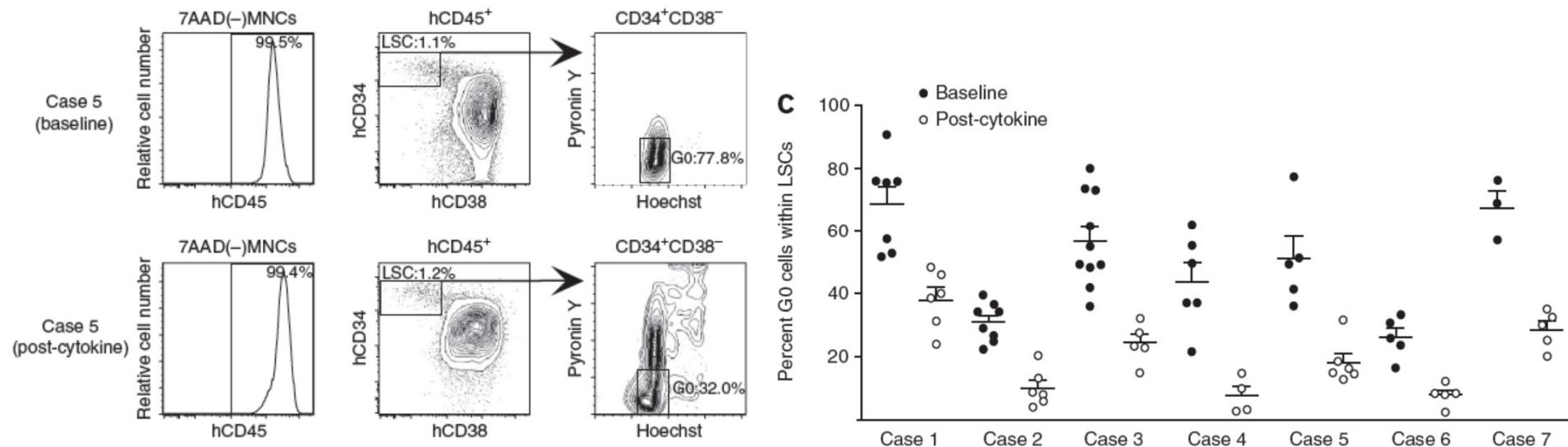


## Intrinsic Resistance of Tumorigenic Breast Cancer Cells to Chemotherapy

Xiaoxian Li, Michael T. Lewis, Jian Huang, Carolina Gutierrez, C. Kent Osborne, Meng-Fen Wu, Susan G. Hilsenbeck, Anne Pavlick, Xiaomei Zhang, Gary C. Chamness, Helen Wong, Jeffrey Rosen, Jenny C. Chang

moreover: CSC show increased resistance to DNA damaging agents (e.g. radiation)

# Therapeutic approaches sensitizing CSCs to standard therapy

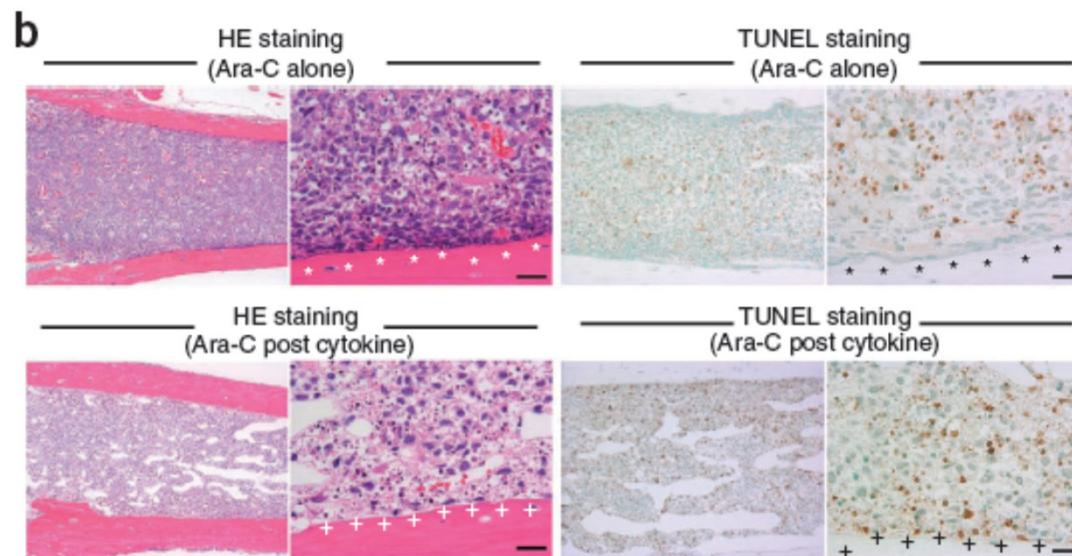
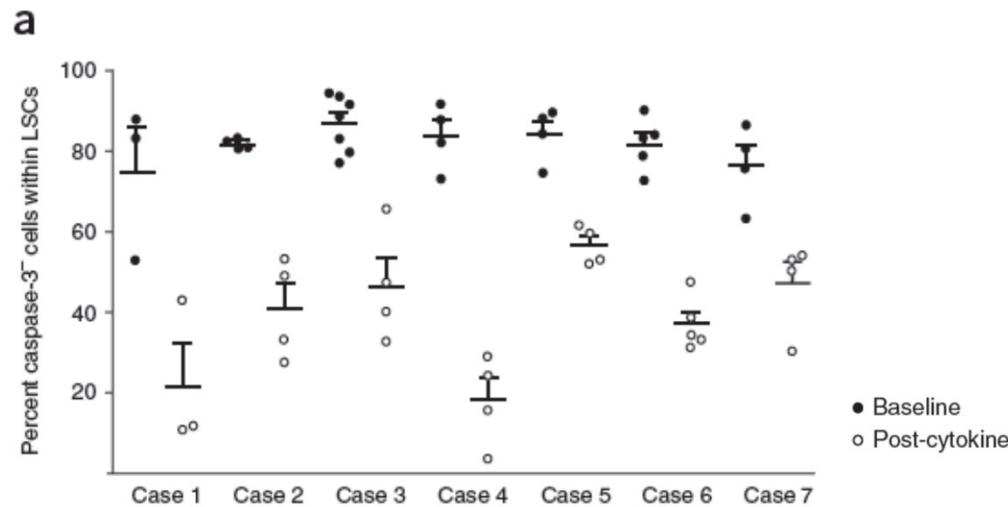


**nature**  
**biotechnology**

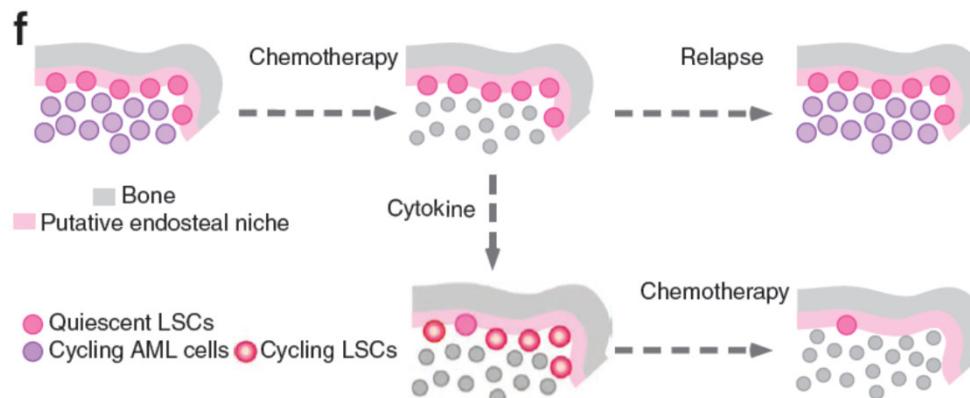
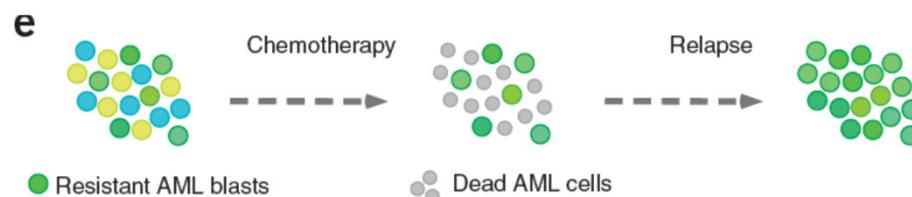
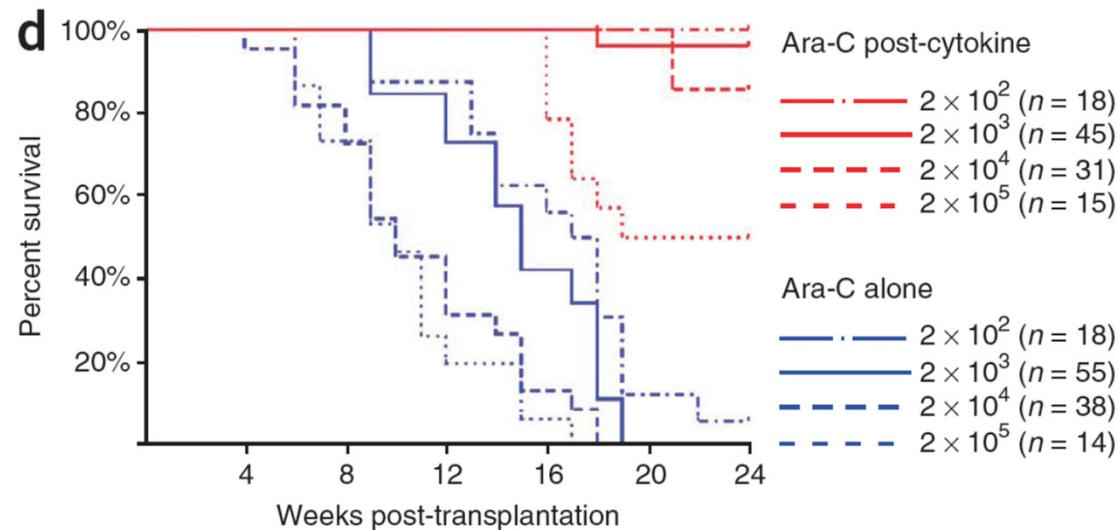
Induction of cell cycle entry eliminates human leukemia stem cells in a mouse model of AML

Yoriko Saito<sup>1</sup>, Naoyuki Uchida<sup>2</sup>, Satoshi Tanaka<sup>3</sup>, Nahoko Suzuki<sup>1</sup>, Mariko Tomizawa-Murasawa<sup>1</sup>, Akiko Sone<sup>1</sup>, Yuho Najima<sup>1</sup>, Shinsuke Takagi<sup>1,2</sup>, Yuki Aoki<sup>1</sup>, Atsushi Wake<sup>2</sup>, Shuichi Taniguchi<sup>2</sup>, Leonard D Shultz<sup>4</sup> & Fumihiko Ishikawa<sup>1</sup>

# Therapeutic approaches sensitizing CSCs to standard therapy

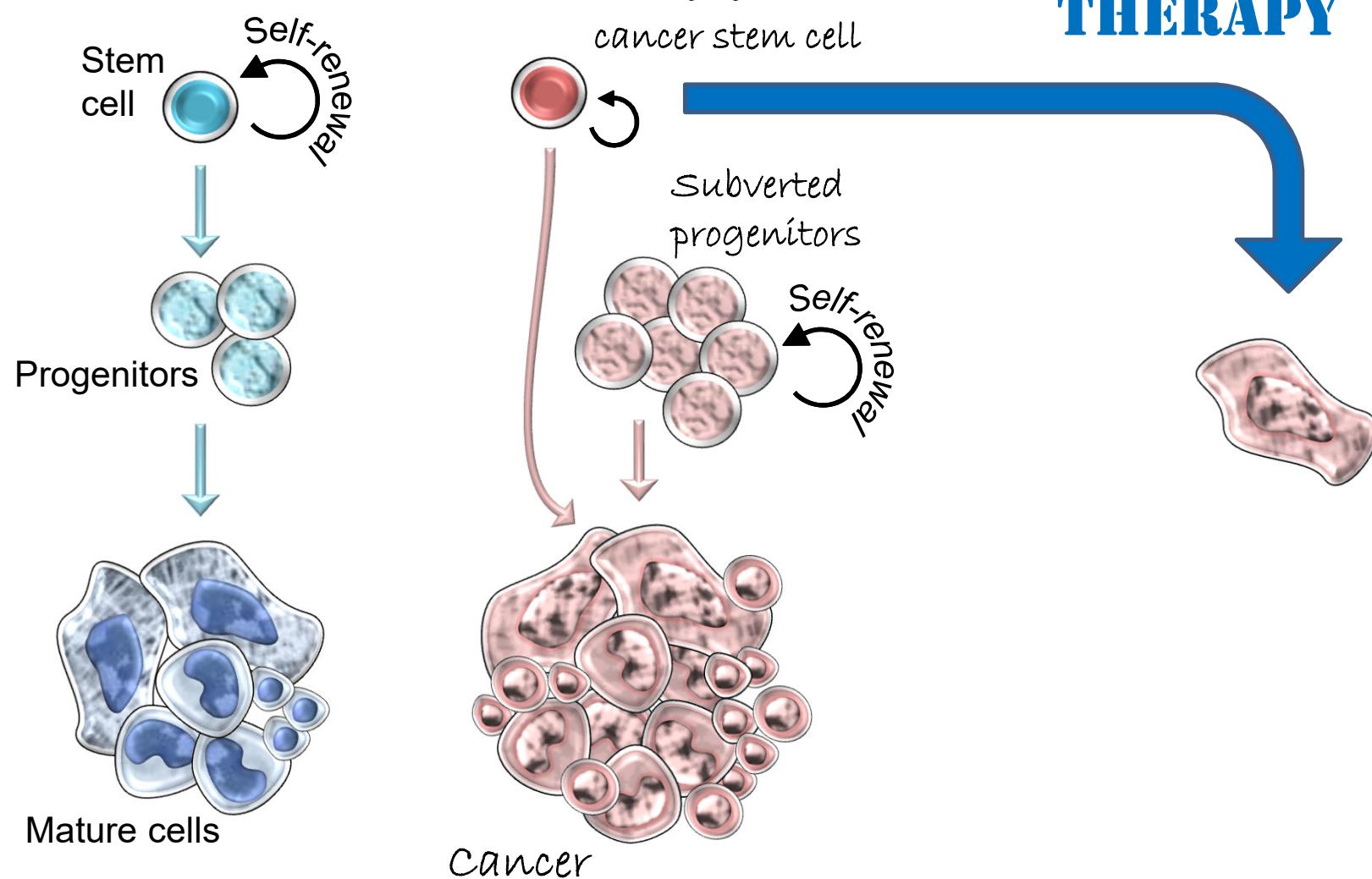


# Therapeutic approaches sensitizing CSCs to standard therapy

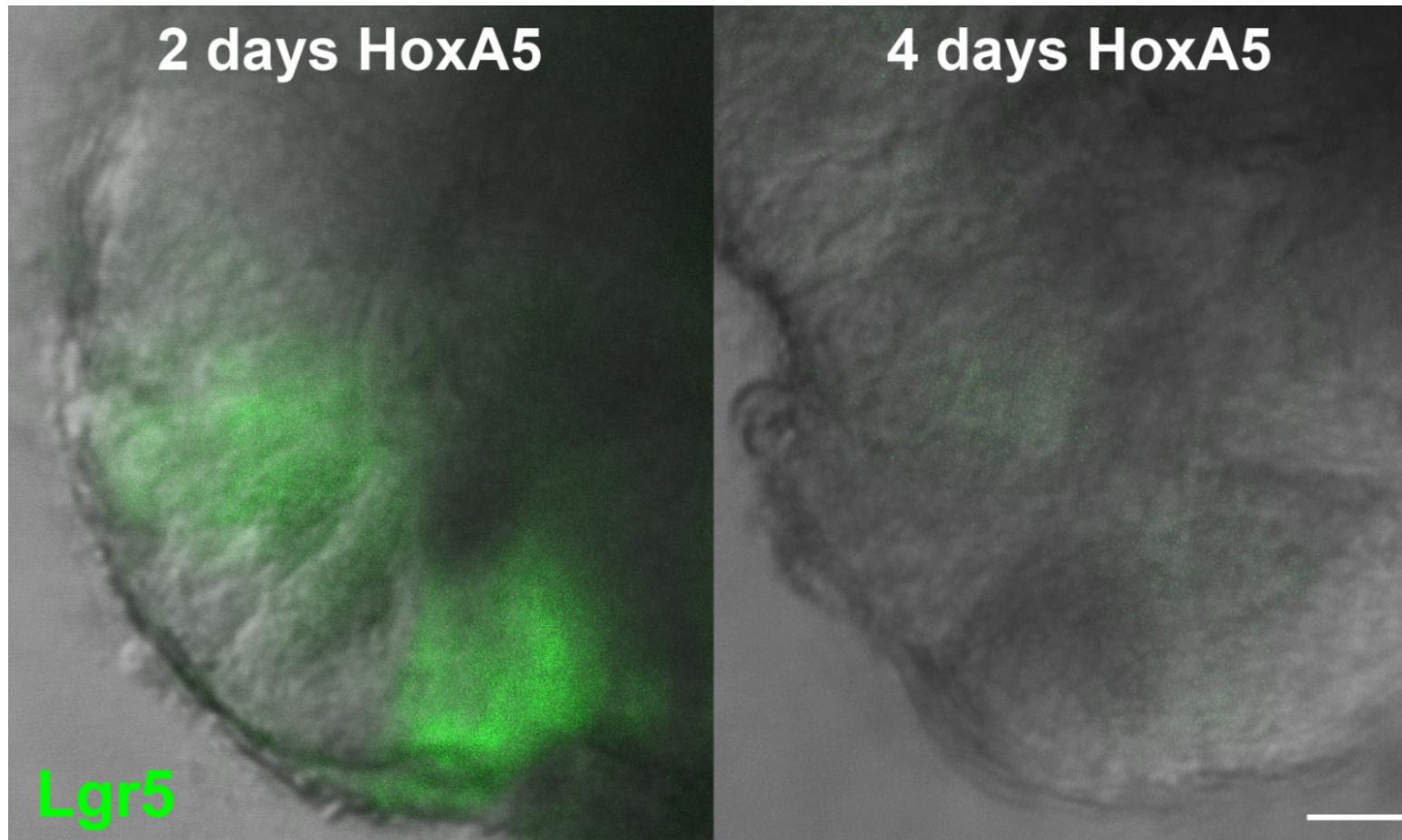
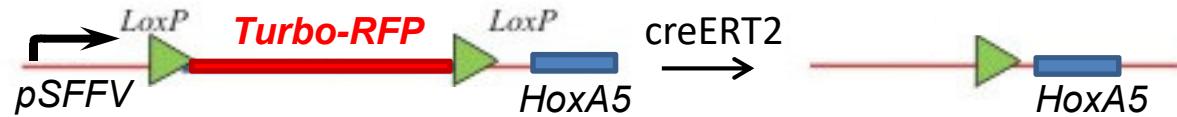


# Stem cells in Cancer

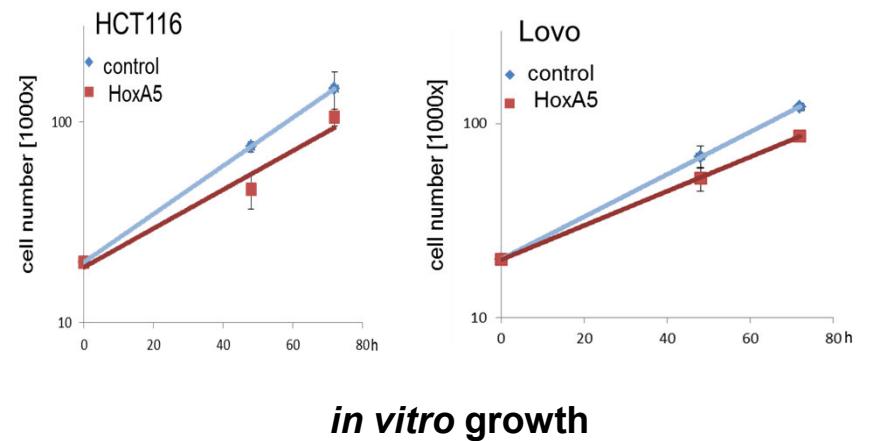
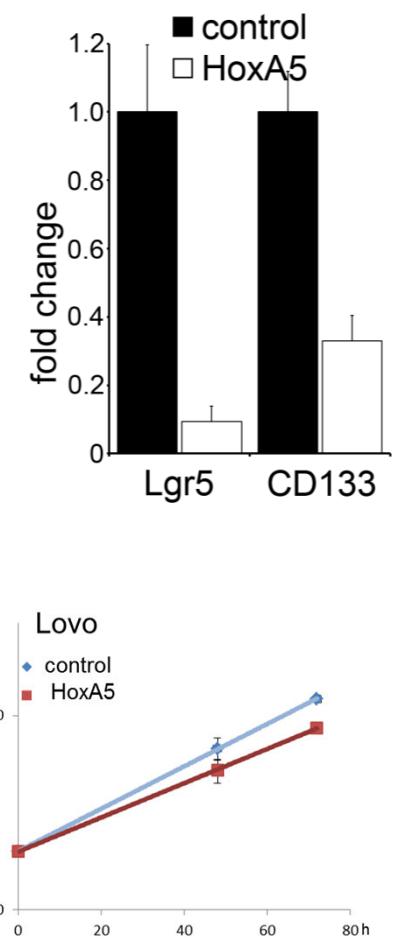
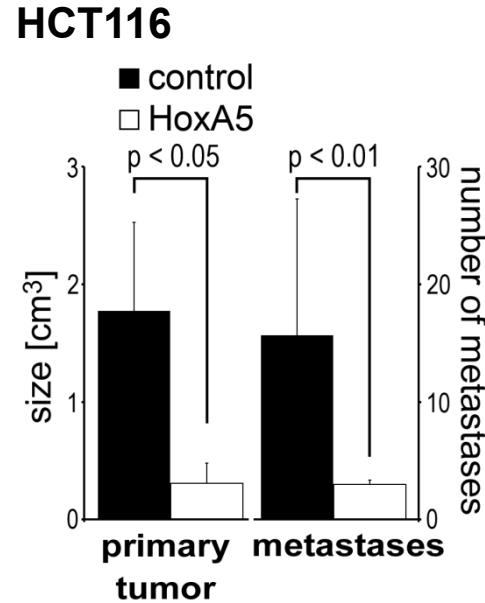
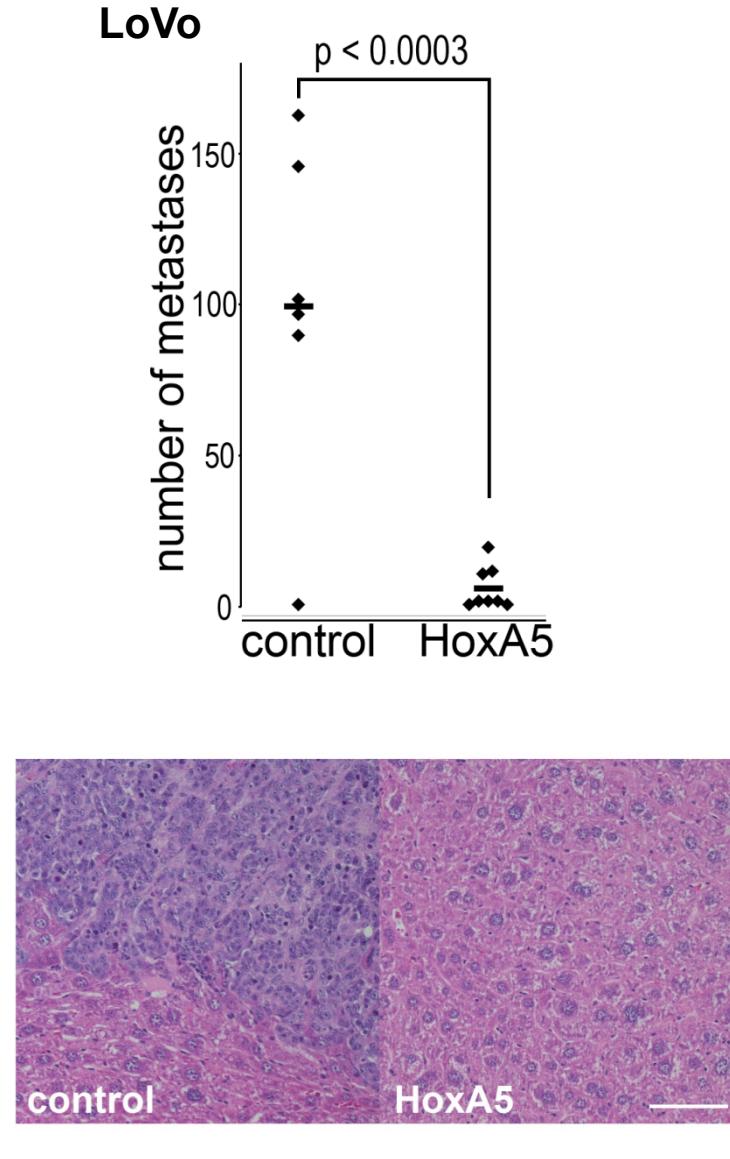
## DIFFERENTIATION THERAPY



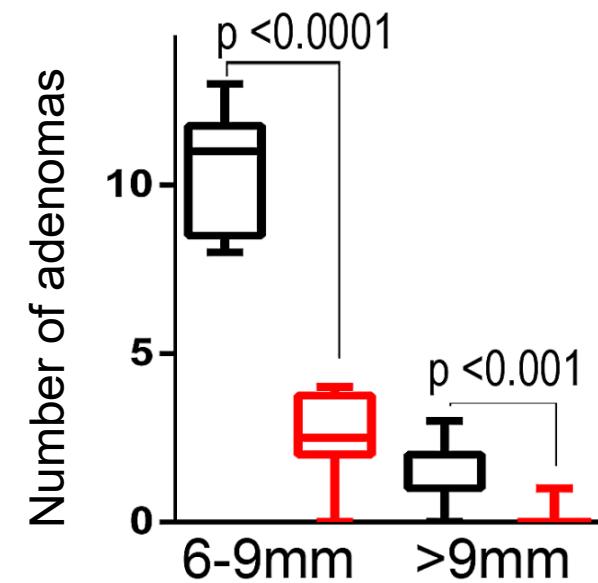
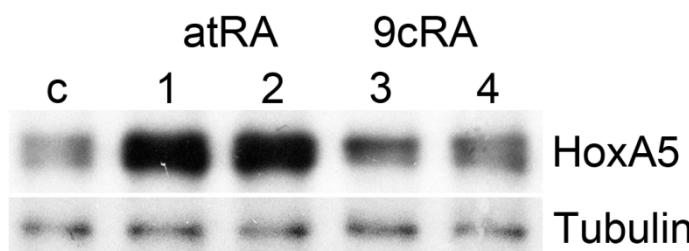
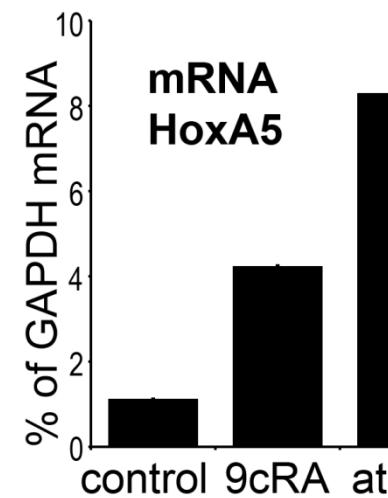
# HoxA5 expression reduces intestinal Lgr5<sup>+</sup> stem cells



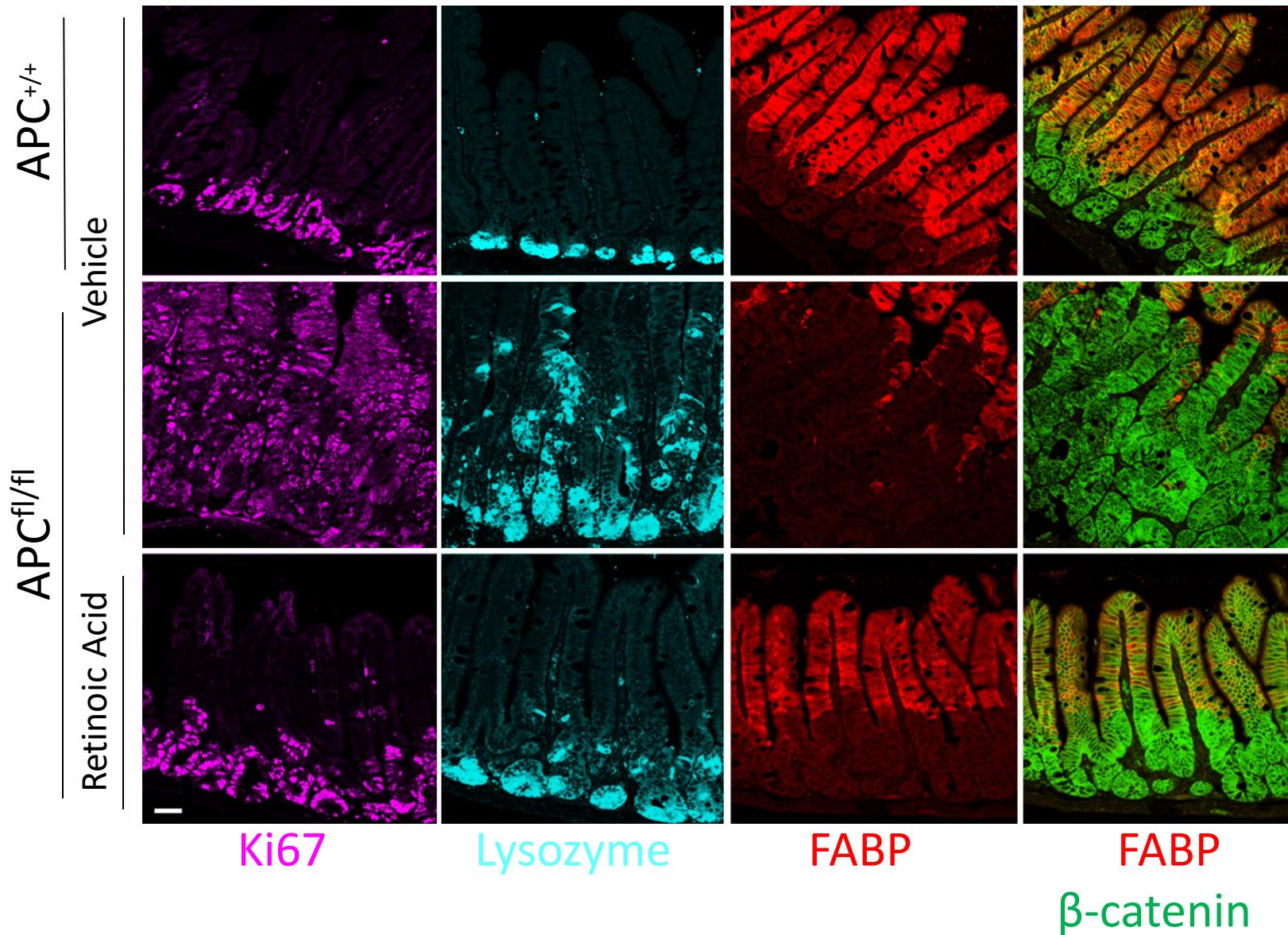
## HoxA5 expression reduces Cancer Stem Cell properties *in vivo*



# Retinoic Acid counteracts tumour initiation driven by Wnt *in vivo*



# atRA Counteracts Tumour Initiation Driven by Wnt *in vivo*



# Retinoid induced differentiation therapy requires HoxA5

